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TRANSMITTAL LETTER TO THE UNITED STATES

International Preliminary Examination Report

ATTORNEY'S DOCKET NUMBER 49100

DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

INTERNATIONAL APPLICATION NO. INTERNATIONAL FILING DATE PRIORITY DATE CLAIMED PCT/EP 99/03889 4 June 1999 5 June 1998 1 March 1999 TITLE OF INVENTION: POLY(ADP-RIBOSE)POLYMERASE-GENE APPLICANT(S) FOR DO/EO/US Michael KOCK, Thomas HOEGER, Burkhard KROEGER, Bernd OTTERBACH Wilfried LUBISCH, Hans-Georg LEMAIRE Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information: 1. /X/ This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 2.11 This express request to begin national examination procedures (35 U.S.C.371(f)) at any time rather than delay examination until 3. /X/ the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1). A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. 4. /x / 5. /X/ A copy of the International Application as filed (35 U.S.C. 371(c)(2)). a./X/ is transmitted herewith (required only if not transmitted by the International Bureau). has been transmitted by the International Bureau. is not required, as the application was filed in the United States Receiving Office (RO/US0). 6. /X/ A translation of the International Application into English (35 U.S.C. 371(c)(2)). 7. /X / Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)). are transmitted herewith (required only if not transmitted by the International Bureau). a./X / b.// have been transmitted by the International Bureau. c.// have not been made; however, the time limit for making such amendments has NOT expired. d.// have not been made and will not be made. 8. /X / A translation of the amendments to the claims under PCT Article 19(35 U.S.C. 371(c)(3)). An oath or declaration of the inventor(s)(35 U.S.C. 171(c)(4)). 9. /X / 10.// A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). Items 11, to 16, below concern other document(s) or information included: 11// An Information Disclosure Statement under 37 CFR 1.97 and 1.98. 12./X / An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 13./X / A FIRST preliminary amendment. 11 A SECOND or SUBSEQUENT preliminary amendment. 14// A substitute specification. 15.// A change of power of attorney and/or address letter. 16./x/ Other items or information. International Search Report

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In re Application of

Box Sequence

KOCK et al.

Serial No. 09/701,586

Filed: November 30, 2000

POLY(ADP-RIBOSE) POLYMERASE-GENE For:

> I hereby certify that this correspondence is being deposited with the United States Postal Service as first class mail in an envelope addressed to: Commissioner of Patents and Trademarks, Washington, D.C. 20231, April 23, 2002

Herbert B. Keil Date of Deposit

Person Making Deposit

Signature April 23, 2002

Date of Signature

Honorable Commissioner of Patents and Trademarks Washington, D.C. 20231

PRELIMINARY AMENDMENT <u>AND</u> RESPONSE TO NOTIFICATION OF DEFECTIVE RESPONSE

Sir:

(1)

In response to the Notification of Defective Response, mailed April 11, 2002, applicants respectfully request entry of the following amendments, in accordance with 37 CFR §1.115.

KOCK et al., Serial No. 09/701,586

CLEAN VERSION OF AMENDMENTS

IN THE SPECIFICATION

113

Please replace the sequence listing on pages 48 to 82 of the specification with the substitute sequence listing appended hereto, numbered pages 1 to 36.

KOCK et al., Serial No. 09/701,586

REMARKS

In response to the Notice of Defective Response, a copy of the substitute sequence listing in computer readable form is attached hereto. The content of the paper copy of the sequence listing and the copy of the sequence listing in computer readable form is the same, and includes no new matter.

It is believed that by submitting the present amendment and the sequence listing diskette, the application now fully complies with the requirements of 37 CFR §§ 1.821-1.825. Applicants respectfully solicit issuance of the patent.

Please charge any shortage in fees due in connection with the filing of this paper, including Extension of Time fees to Deposit Account No. 11-0345. Please credit any excess fees to such deposit account.

Respectfully submitted, KEIL & WEINKAUF

David C. Liechty Reg. No. 48,692

1101 Connecticut Ave., N.W. Washington, D.C. 20036 (202)659-0100

DCL/kas

529 Rec'd PCT/PTC 3 0 NOV 2000

IN THE UNITED STA	TES PATENT AND TRADEMARK OFFICE
In re the Application of KOCK et al.	BOX PCT
International Application PCT/EP 99/03889)))
Filed: June 4, 1999))
For: POLY(ADP-RIBOSE)POLYMER) ASE-GENE

PRELIMINARY AMENDMENT

Honorable Commissioner of Patents and Trademarks Washington, D.C. 20231

Sir:

Prior to examination, kindly amend the above-identified application as follows: IN THE CLAIMS

3. A PARP homolog as claimed in <u>claim 1</u> [either of the preceding claims], comprising at least another one of the following part-sequence motifs:

LX₉NX₂YX₂QLLX(D/E)X_{10/11}WGRVG, AX₃FXKX₄KTXNXWX₅FX₃PXK, QXL(I/L)X₂IX₉MX₁₀PLGKLX₃QIX₆L, FYTXIPHXFGX₃PP; and KX₃LX₂LXDIEXAX₂L,

in which the X radicals are, independently of one another, any amino acid.

4. A PARP homolog as claimed in <u>claim 1</u> [any of the preceding claims], selected from human PARP homologs, which has the amino acid sequence shown in SEQ ID NO: 2 (human PARP2) or SEQ ID NO: 4 or 6 (human PARP3 type 1 or 2); or murine PARP homologs which have the amino acid sequence shown in SEQ ID NO:8 (mouse PARP long form) or SEQ ID No:10 (mouse PARP short form); and the functional equivalents thereof.

- 5. A binding partner for PARP homologs as claimed in <u>claim 1</u> [any of the preceding claims], selected from
- a) antibodies and fragments thereof,
- b) protein-like compounds which interact with a part-sequence of the protein, and
- c) low molecular weight effectors which modulate the catalytic PARP activity or another biological function of a PARP molecule.
- 6. A nucleic acid comprising
- a) a nucleotide sequence coding for at least one PARP homolog as claimed in <u>claim 1</u> [any of claims 1 to 4], or the complementary nucleotide sequence thereof;
- b) a nucleotide sequence which hybridizes with a sequence as specified in a) under stringent conditions; or
- c) nucleotide sequences which are derived from the nucleotide sequences defined in
- a) and b) through the degeneracy of the genetic code.
- 8. An expression cassette comprising, under the genetic control of at least one regulatory nucleotide sequence, at least one nucleotide sequence as claimed in <u>claim 6</u> [either of claims 6 and 7].
- 12. A PARP-deficient mammal or PARP-deficient eukaryotic cell, in which functional expression of at least one gene which codes for a PARP homolog as claimed in <u>claim 1</u> [any of claims 1 to 4] is inhibited.
- 13. An in vitro detection method for PARP inhibitors, which comprises
 - incubating an unsupported or supported polyADP-ribosylatable target with a reaction mixture comprising
 - a1) a PARP homolog as claimed in claim 1 [any of claims 1 to 4],
 - a2) a PARP activator; and
 - a PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected;
 - b) carrying out the polyADP ribosylation reaction; and

- c) determining the polyADP ribosylation of the target qualitatively or quantitatively.
 - 15. A method as claimed in <u>claim 13</u> [either of claims 13 and 14], wherein the polyADP-ribosylatable target is a histone protein.
- A method as claimed in <u>claim 13</u> [any of claims 13 to 15], wherein the PARP activator is activated DNA.
- 17. A method as claimed in <u>claim 13</u> [any of claims 13 to 16], wherein the polyADP ribosylation reaction is started by adding NAD⁺.
- 18. A method as claimed in <u>claim 13</u> [any of claims 13 to 17], wherein the polyADP ribosylation of the supported target is determined using anti-poly(ADP-ribose) antibodies.
- 19. A method as claimed in <u>claim 13</u> [any of claims 13 to 17], wherein the unsupported target is labeled with an acceptor fluorophore.
 - 21. A method as claimed in <u>claim 19</u> [either of claims 19 and 20], wherein the target is biotinylated histone, and the acceptor fluorophore is coupled thereto via avidin or streptavidin.
- 22. A method as claimed in <u>claim 20</u> [either of claims 20 and 21], wherein the anti-poly(ADP-ribose) antibody carries a europium cryptate as donor fluorophore.
- 23. An in vitro screening method for binding partners for a PARP molecule, which comprises
- a1) immobilizing at least one PARP homolog as claimed in <u>claim 1</u> [any of claims 1 to 4] on a support;
- b1) contacting the immobilized PARP homolog with an analyte in which at least one binding partner is suspected; and

c1) determining, where appropriate after an incubation period, analyte constituents bound to the immobilized PARP homolog;

or

- a2) immobilizing on a support an analyte which comprises at least one possible binding partner for a PARP molecule;
- b2) contacting the immobilized analyte with at least one PARP homolog as claimed in claim 1 [any of claims 1 to 4] for which a binding partner is sought; and
- c2) examining the immobilized analyte, where appropriate after an incubation period, for binding of the PARP homolog.
- 24. A method for the qualitative or quantitative determination of nucleic acids encoding a PARP homolog as claimed in <u>claim 1</u> [any of claims 1 to 4], which comprises
 - a) incubating a biological sample with a defined amount of an exogenous nucleic acid [as claimed in either of claims 6 and 7], hybridizing under stringent conditions, determining the hybridizing nucleic acids and, where appropriate, comparing with a standard; or
 - b) incubating a biological sample with a pair of oligonucleotide primers with specificity for a PARP homolog-encoding nucleic acid, amplifying the nucleic acid, determining the amplification product and, where appropriate, comparing with a standard.
- 25. A method for the qualitative or quantitative determination of a PARP homolog as claimed in claim 1 [any of claims 1 to 4], which comprises
 - a) incubating a biological sample with a binding partner specific for a

- PARP homolog,
- b) detecting the binding partner/PARP complex and, where appropriate,
- c) comparing the result with a standard.
 - 27. A method as claimed in <u>claim 24</u> [any of claims 24 to 26] for diagnosing energy deficit-mediated illnesses.
- 28. A method for determining the efficacy of PARP effectors, which comprises
- incubating a PARP homolog as claimed in <u>claim 1</u> [any of claims 1 to 4] with an analyte which comprises an effector of a physiological or pathological PARP activity; removing the effector again where appropriate; and
- b) determining the activity of the PARP homolog, where appropriate after adding substrates or cosubstrates.
- 29. A gene therapy composition, which comprises in a vehicle acceptable for gene therapy a nucleic acid construct which
 - a) comprises an antisense nucleic acid against a coding nucleic acid as claimed in claim 6 [either of claims 6 and 7]; or
 - b) a ribozyme against a nucleic acid as claimed in <u>claim 6</u> [either of claims
 6 and 7]; or
 - c) codes for a specific PARP inhibitor.
- 30. A pharmaceutical composition comprising, in a pharmaceutically acceptable vehicle, at least one PARP protein as claimed in <u>claim 1</u> [any of claims 1 to 4], at least one PARP binding partner [as claimed in claim 5] or at least one coding nucleotide sequence [as claimed in claim 6 or 7].

REMARKS

The claims have been amended to eliminate multiple dependency and to put them in better form for U.S. filing. No new matter is included. A clean copy of the claims is attached.

Favorable action is solicited.

Respectfully submitted,

KEIL & WEINKAUF

Herbert B. Keil

Reg. No. 18,967

1101 Connecticut Ave., N.W. Washington, D.C. 20036 (202)659-0100

CLEAN COPY OF THE CLAIMS

- 1. A poly(ADP-ribose) polymerase (PARP) homolog which has an amino acid sequence which has
 - a) a functional NAD⁺ binding domain and
 - b) no zinc finger sequence motif of the general formula

CX2CXmHX2C

in which
m is an integral value from 28 or 30, and the X radicals are,
independently of one another, any amino acid;
and the functional equivalents thereof.

2. A PARP homolog as claimed in claim 1, wherein the functional NAD⁺ binding domain comprises one of the following general sequence motifs:

PX_n(S/T)GX₃GKGIYFA, (S/T)XGLR(I/V)XPX_n(S/T)GX₃GKGIYFA or LLWHG(S/T)X₇IL(S/T)XGLR(I/V)XPX_n(S/T)GX₃GKGIYFAX₃SKSAXY

in which

n is an integral value from 1 to 5, and the X radicals are, independently of one another, any amino acid.

3. A PARP homolog as claimed in claim 1, comprising at least another one of the following part-sequence motifs:

LX₉NX₂YX₂QLLX(D/E)X_{10/11}WGRVG, AX₃FXKX₄KTXNXWX₅FX₃PXK,

QXL(I/L) $X_2IX_9MX_{10}PLGKLX_3QIX_6L$, FYTXIPHXFGX₃PP; and KX₃LX₂LXDIEXAX₂L,

in which the X radicals are, independently of one another, any amino acid.

- 4. A PARP homolog as claimed in claim 1, selected from human PARP homologs, which has the amino acid sequence shown in SEQ ID NO: 2 (human PARP2) or SEQ ID NO: 4 or 6 (human PARP3 type 1 or 2); or murine PARP homologs which have the amino acid sequence shown in SEQ ID NO:8 (mouse PARP long form) or SEQ ID No:10 (mouse PARP short form); and the functional equivalents thereof.
- 5. A binding partner for PARP homologs as claimed in claim 1, selected from
 - a) antibodies and fragments thereof,
 - b) protein-like compounds which interact with a part-sequence of the protein, and
 - c) low molecular weight effectors which modulate the catalytic PARP activity or another biological function of a PARP molecule.
- 6. A nucleic acid comprising
 - a) a nucleotide sequence coding for at least one PARP homolog as claimed in claim 1, or the complementary nucleotide sequence thereof;
 - a nucleotide sequence which hybridizes with a sequence as specified in
 a) under stringent conditions; or
 - c) nucleotide sequences which are derived from the nucleotide sequences defined in a) and b) through the degeneracy of the genetic code.
- 7. A nucleic acid as claimed in claim 6, comprising
 - a) nucleotides +3 to +1715 shown in SEQ ID NO:1;

- b) nucleotides +242 to +1843 shown in SEQ ID NO:3;
- c) nucleotides +221 to +1843 shown in SEQ ID NO:5;
- d) nucleotides +112 to +1710 shown in SEQ ID NO:7; or
- e) nucleotides +1 to +1584 shown in SEQ ID NO:9.
- 8. An expression cassette comprising, under the genetic control of at least one regulatory nucleotide sequence, at least one nucleotide sequence as claimed in claim 6.
- 9. A recombinant vector comprising at least one expression cassette as claimed in claim 8.
- 10. A recombinant microorganism comprising at least one recombinant vector as claimed in claim 9.
- 11. A transgenic mammal comprising a vector as claimed in claim 9.
- A PARP-deficient mammal or PARP-deficient eukaryotic cell, in which functional expression of at least one gene which codes for a PARP homolog as claimed in claim 1 is inhibited.
- 13. An in vitro detection method for PARP inhibitors, which comprises
 - a) incubating an unsupported or supported polyADP-ribosylatable target with a reaction mixture comprising
 - a1) a PARP homolog as claimed in claim 1,
 - a2) a PARP activator; and
 - a3) a PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected;
 - b) carrying out the polyADP ribosylation reaction; and
 - c) determining the polyADP ribosylation of the target qualitatively or

quantitatively.

- 14. A method as claimed in claim 13, wherein the PARP homolog is preincubated with the PARP activator and the PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected, before the polyADP ribosylation reaction is carried out.
- 15. A method as claimed in claim 13, wherein the polyADP-ribosylatable target is a histone protein.
- 16. A method as claimed in claim 13, wherein the PARP activator is activated DNA.
- 17. A method as claimed in claim 13, wherein the polyADP ribosylation reaction is started by adding NAD⁺.
- 18. A method as claimed in claim 13, wherein the polyADP ribosylation of the supported target is determined using anti-poly(ADP-ribose) antibodies.
- 19. A method as claimed in claim 13, wherein the unsupported target is labeled with an acceptor fluorophore.
- 20. A method as claimed in claim 19, wherein the polyADP ribosylation of the unsupported target is determined using anti-poly(ADP-ribose) antibody which is labeled with a donor fluorophore which is able to transfer energy to the acceptor fluorophore.
- 21. A method as claimed in claim 19, wherein the target is biotinylated histone, and the acceptor fluorophore is coupled thereto via avidin or streptavidin.
- 22. A method as claimed in claim 20, wherein the anti-poly(ADP-ribose) antibody

carries a europium cryptate as donor fluorophore.

- 23. An in vitro screening method for binding partners for a PARP molecule, which comprises
 - a1) immobilizing at least one PARP homolog as claimed in claim 1 on a support;
 - b1) contacting the immobilized PARP homolog with an analyte in which at least one binding partner is suspected; and
 - c1) determining, where appropriate after an incubation period, analyte constituents bound to the immobilized PARP homolog;

or

- a2) immobilizing on a support an analyte which comprises at least one possible binding partner for a PARP molecule;
- b2) contacting the immobilized analyte with at least one PARP homolog for which a binding partner is sought; and
- c2) examining the immobilized analyte, where appropriate after an incubation period, for binding of the PARP homolog.
- 24. A method for the qualitative or quantitative determination of nucleic acids encoding a PARP homolog as claimed in claim 1, which comprises
 - incubating a biological sample with a defined amount of an exogenous nucleic acid, hybridizing under stringent conditions, determining the hybridizing nucleic acids and, where appropriate, comparing with a standard; or
 - b) incubating a biological sample with a pair of oligonucleotide primers with specificity for a PARP homolog-encoding nucleic acid, amplifying the nucleic acid, determining the amplification product and, where appropriate, comparing with a standard.

- 25. A method for the qualitative or quantitative determination of a PARP homolog as claimed in claim 1, which comprises
 - incubating a biological sample with a binding partner specific for a PARP homolog,
 - b) detecting the binding partner/PARP complex and, where appropriate,
 - c) comparing the result with a standard.
- 26. A method as claimed in claim 25, wherein the binding partner is an antibody or a binding fragment thereof, which carries a detectable label where appropriate.
- 27. A method as claimed in claim 24 for diagnosing energy deficit-mediated illnesses.
- 28. A method for determining the efficacy of PARP effectors, which comprises
 - incubating a PARP homolog as claimed in claim 1 with an analyte which comprises an effector of a physiological or pathological PARP activity;
 removing the effector again where appropriate; and
 - b) determining the activity of the PARP homolog, where appropriate after adding substrates or cosubstrates.
- 29. A gene therapy composition, which comprises in a vehicle acceptable for gene therapy a nucleic acid construct which
 - a) comprises an antisense nucleic acid against a coding nucleic acid as claimed in claim 6; or
 - b) a ribozyme against a nucleic acid as claimed in claim 6; or
 - c) codes for a specific PARP inhibitor.
- 30. A pharmaceutical composition comprising, in a pharmaceutically acceptable vehicle, at least one PARP protein as claimed in claim 1, at least one PARP binding partner or at least one coding nucleotide sequence

- 31. The use of low molecular weight PARP binding partners as claimed in claim 5 for the diagnosis or therapy of pathological states in the development and/or progress of which at least one PARP protein, or a polypeptide derived therefrom, is involved.
- 32. The use of low molecular weight PARP binding partners as claimed in claim 5 for the diagnosis or therapy of pathological states mediated by an energy deficit.

7/PRTS 09/701586 529 Rec'd PCT/PTC 30 NOV 2000

Novel poly(ADP-ribose) polymerase genes

The present invention relates to novel poly(ADP-ribose) polyme5 rase (PARP) genes and to the proteins derived therefrom;
antibodies with specificity for the novel proteins;
pharmaceutical and gene therapy compositions which comprise
products according to the invention; methods for the analytical
determination of the proteins and nucleic acids according to the
10 invention; methods for identifying effectors or binding partners
of the proteins according to the invention; methods for
determining the activity of such effectors and use thereof for
the diagnosis or therapy of pathological states.

- 15 In 1966, Chambon and co-workers discovered a 116 kD enzyme which was characterized in detail in subsequent years and is now called PARP (EC 2.4.2.30) (poly(adenosine-5'-diphosphoribose) polymerase), PARS (poly(adenosine-5'-diphosphoribose) synthase) or ADPRT (adenosine-5'-diphosphoribose transferase). In the plant
- 20 kingdom (Arabidopsis thaliana) a 72kD (637 amino acids) PARP was found in 1995 (Lepiniec L. et al., FEBS Lett 1995; 364(2): 103-8). It was not clear whether this shorter form of PARP is a plant-specific individuality or an artefact ("splice" variant or the like). The 116 kD PARP enzyme has to date been unique in
- 25 animals and in man in its activity, which is described below. It is referred to as PARP1 below to avoid ambiguity.

The primary physiological function of PARP 1 appears to be its involvement in a complex repair mechanism which cells have

- 30 developed to repair DNA strand breaks. The primary cellular response to a DNA strand break appears moreover to consist of PARP1-catalyzed synthesis of poly(ADP-ribose) from NAD+ (cf. De Murcia, G. et al. (1994) TIBS, 19, 172).
- 35 PARP 1 has a modular molecular structure. Three main functional elements have been identified to date: an N-terminal 46 kD DNA binding domain; a central 22 kD automodification domain to which poly(ADP-ribose) becomes attached, with the PARP 1 enzyme activity decreasing with increasing elongation; and a C-terminal
- 40 54 kD NAD+ binding domain. A leucine zipper region has been found within the automodification domain, indicating possible protein-protein interactions, only in the PARP from Drosophila. All PARPs known to date are presumably active as homodimers.
- 45 The high degree of organization of the molecule is reflected in the strong conservation of the amino acid sequence. Thus, 62% conservation of the amino acid sequence has been found for PARP 1

from humans, mice, cattle and chickens. There are greater structural differences from the PARP from Drosophila. The individual domains themselves in turn have clusters of increased conservation. Thus, the DNA binding region contains two so-called zinc fingers as subdomains (comprising motifs of the type CX2CX28/30HX2C), which are involved in the Zn2+-dependent recognition of DNA single strand breaks or single-stranded DNA overhangs (e.g. at the chromosome ends, the telomeres). The C-terminal catalytic domain comprises a block of about 50 amino acids (residues 859-908), which is about 100% conserved among vertebrates (PARP "signature"). This block binds the natural substrate NAD+ and thus governs the synthesis of poly(ADP-ribose) (cf. de Murcia, loc.cit.). The GX3GKG motif in particular is characteristic of PARPs in this block.

15

The beneficial function described above contrasts with a pathological one in numerous diseases (stroke, myocardial infarct, sepsis etc.). PARP is involved in cell death resulting from ischemia of the brain (Choi, D.W., (1997) Nature Medicine,

- 20 3, 10, 1073), of the myocardium (Zingarelli, B., et al (1997), Cardiovascular Research, 36, 205) and of the eye (Lam, T.T. (1997), Res. Comm. in Molecular Pathology and Pharmacology, 95, 3, 241). PARP activation induced by inflammatory mediators has also been observed in septic shock (Szabo, C., et al. (1997),
- 25 Journal of Clinical Investigation, 100, 3, 723). In these cases, activation of PARP is accompanied by extensive consumption of NAD+. Since four moles of ATP are consumed for the biosynthesis of one mole of NAD+, the cellular energy supply decreases drasticallly. The consequence is cell death.

30

PARP1 inhibitors described in the abovementioned specialist literature are nicotinamide and 3-aminobenzamide. 3,4-Di-hydro-5-[4-(1-piperidinyl)butoxy]-1(2H)-isoquinolone is disclosed by Takahashi, K., et al (1997), Journal of Cerebral Blood Flow and Metabolism 17, 1137. Further inhibitors are described, for example, in Banasik, M., et al. (1992) J. Biol. Chem., 267, 3, 1569 and Griffin, R.J., et al. (1995), Anti-Cancer Drug Design,

10, 507.

40 High molecular weight binding partners described for human PARP1 include the base excision repair (BER) protein XRCC1 (X-ray repair cross-complementing 1) which binds via a zinc finger motif and a BRCT (BRCA1 C-terminus) module (amino acids 372-524) (Masson, M., et al., (1998) Molecular and Cellular Biology, 18,6,45 3563).

It is an object of the present invention, because of the diverse physiological and pathological functions of PARP, to provide novel PARP homologs. The reason for this is that the provision of homologous PARPs would be particularly important for developing novel targets for drugs, and novel drugs, in order to improve diagnosis and/or therapy of pathological states in which PARP, PARP homologs or substances derived therefrom are involved.

We have found that this object is achieved by providing PARP 10 homologs, preferably derived from human and non-human mammals, having an amino acid sequence which has

- a functional NAD+ binding domain, i.e. a PARP "signature" sequence having the characteristic GX₃GKG motif;
 and
- especially in the N-terminal sequence region, i.e. in the region of the first 200, such as, for example, in the region of the first 100, N-terminal amino acids, no PARP zinc finger sequence motifs of the general formula

 $CX_2CX_mHX_2C$

- 20 in which
 - m is an integral value from 28 or 30, and the X radicals are, independently of one another, any amino acid; and the functional equivalents thereof.
- 25 Since the PARP molecules according to the invention represent in particular functional homologs, they naturally also have a poly(ADP-ribose)-synthesizing activity. The NAD binding domain essentially corresponds to this activity and is localized to the C terminus.
- 30

Thus an essential characteristic of the PARPs according to the invention is the presence of a functional NAD+ binding domain (PARP signature) which is located in the C-terminal region of the amino acid sequence (i.e. approximately in the region of the last

- 35 400, such as, for example, the last 350 or 300, C-terminal amino acids), in combination with an N-terminal sequence having no zinc finger motifs. Since the zinc finger motifs in known PARPs presumably contribute to recognition of the DNA breakages, it is to be assumed that the proteins according to the invention do not
- 40 interact with DNA or do so in another way. It has been demonstrated by appropriate biochemical tests that the PARP2 according to the invention can be activated by 'activated DNA' (i.e. DNA after limited DNaseI digestion). It can be concluded from this further that the PARP2 according to the invention has
- 45 DNA binding properties. However, the mechanism of the DNA binding and enzyme activation differs between the PARPs according to the invention and PARP1. Its DNA binding and enzyme activation is, as

mentioned, mediated by a characteristic zinc finger motif. No such motifs are present in the PARPs according to the invention. Presumably these properties are mediated by positively charged amino acids in the N-terminal region of the PARPs according to 5 the invention. Since the 'activated DNA' (i.e. for example DNA after limited treatment with DNaseI) has a large number of defects (single strand breaks, single strand gaps, single-stranded overhangs, double strand breaks etc.), it is possible that although PARPI and the PARPs according to the invention are activated by the same 'activated DNA', it is by a different subpopulation of defects (e.g. single strand gaps instead of single strand breaks).

The functional NAD+ binding domain (i.e. catalytic domain) binds

15 the substrate for poly-(ADP-ribose) synthesis. Consistent with
known PARPs, the sequence motif GX1X2X3GKG, in which G is glycine,
K is lysine, and X1, X2 and X3 are, independently of one another,
any amino acid, is present in particular. However, as shown,
surprisingly, by comparison of the amino acid sequences of the

20 NAD+ binding domains of PARP molecules according to the invention
with previously disclosed human PARP1, the sequences according to
the invention differ markedly from the known sequence for the NAD+
binding domain.

25 A group of PARP molecules which is preferred according to the invention preferably has the following general sequence motif in the catalytic domain in common:

PX_n(S/T)GX₃GKGIYFA (SEQ ID NO:11), in particular

(S/T)XGLR(I/V)XPX_n(S/T)GX₃GKGIYFA (SEQ ID NO:12),

preferably

LLWHG(S/T)X₇IL(S/T)XGLR(I/V)XPX_n(S/T)GX₃GKGIYFAX₃SKSAXY

(SEQ ID NO:13)

35 in which (S/T) describes the alternative occupation of this sequence position by S or T, (I/V) describes the alternative occupation of this sequence position by I or V, and n is an integral value from 1 to 5, and the X radicals are, independently of one another, any amino acid. The last motif is also referred to as the "PARP signature" motif.

The automodification domain is preferably likewise present in the PARPs according to the invention. It can be located, for example, in the region from about 100 to 200 amino acids in front of the 45 N-terminal end of the NAD+ binding domain.

PARP homologs according to the invention may additionally comprise, N-terminally of the NAD+ binding domain (i.e. about 30 to about 80 amino acids closer to the N terminus), a leucine zipper-like sequence motif of the general formula

 $(L/V)X_6LX_6LX_6L$ (SEQ ID NO:14)

in which

5

(L/V) represents the alternative occupation of this sequence position by L or V, and the X radicals are, independently of one another, any amino acid. The leucine zipper motifs observed 10 according to the invention differ distinctly in position from those described for PARP from Drosophila. Leucine zippers may lead to homodimers (two PARP molecules) or heterodimers (one PARP molecule with a binding partner differing therefrom).

15 The PARP homologs according to the invention preferably additionally comprise, N-terminally of the abovementioned leucine zipper-like sequence motifs, i.e. about 10 to 250 amino acid residues closer to the N terminus, at least another one of the following part-sequence motifs:

20

25

LX ₉ NX ₂ YX ₂ QLLX(D/E)X _b WGRVG,	(motif 1; SEQ ID NO:15)
AX3FXKX4KTXNXWX5FX3PXK,	(motif 2; SEQ ID NO:16)
QXL(I/L) $X_2IX_9MX_{10}PLGKLX_3QIX_6L$,	(motif 3; SEQ ID NO:17)
FYTXIPHXFGX3PP,	(motif 4; SEQ ID NO:18)
and	
KX3LX2LXDIEXAX2L	(motif 5; SEQ ID NO:19),

in which (D/E) describes the alternative occupation of this sequence position by D or E, (I/L) describes the alternative

30 occupation of this sequence position by I or L, b is the integral value 10 or 11, and the X radicals are, independently of one another, any amino acid. It is most preferred for these motifs 1 to 5 all to be present in the stated sequence, with motif 1 being closest to the N terminus.

35

The abovementioned PARP signature motif is followed in the proteins according to the invention by at least another one of the following motifs:

```
GX<sub>3</sub>LXEVALG (motif 6; SEQ ID NO:20)

40 GX<sub>2</sub>SX<sub>4</sub>GX<sub>3</sub>PX<sub>a</sub>LXGX<sub>2</sub>V (motif 7; SEQ ID NO:21) and

E(Y/F)X<sub>2</sub>YX<sub>3</sub>QX<sub>4</sub>YLL (motif 8; SEQ ID NO:22)
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in which (Y/F) describes the alternative occupation of this sequence position by Y or F, a is equal to 7 to 9 and X is in 45 each case any amino acid. It is most preferred for the three

C-terminal motifs all to be present and in the stated sequence, with motif 8 being closest to the C terminus.

A preferred PARP structure according to the invention may be 5 described schematically as follows:

Motifs 1 to 5/PARP signature/motifs 6 to 8 or motifs 1 to 5/leucine zipper/PARP signature/motifs 6 to 8

- 10 it being possible for further amino acid residues, such as, for example, up to 40, to be arranged between the individual motifs and for further amino acid residues, such as, for example, up to 80, to be arranged at the N terminus and/or at the C terminus.
- 15 PARP homologs which are particularly preferred according to the invention are the proteins human PARP2, human PARP3, mouse PARP3 and the functional equivalents thereof. The protein referred to as human PARP2 comprises 570 amino acids (cf. SEQ ID NO:2). The protein referred to as human PARP3 possibly exists in two forms.
- 20 Type 1 comprises 533 amino acids (SEQ ID NO:4) and type 2 comprises 540 amino acids (SEQ ID NO:6). The forms may arise through different initiation of translation. The protein referred to as mouse PARP3 exists in two forms which differ from one another by a deletion of 5 amino acids (15 bp). Type 1 comprises
- 25 533 amino acids (SEQ ID NO: 8) and type 2 comprises 528 amino acids (SEQ ID NO:10). The PARP-homologs of the present invention differ in their sequences significantly over said PARP protein of Arabidopsis thaliana (see above). For example, PARP2 and PARP3 do not comprise the plant PARP specific peptide sequence AAVLDQWIPD,
- 30 corresponding to amino acid residues 143 to 152 of the Arabidopsis protein.

The invention further relates to the binding partners for the PARP homologs according to the invention. These binding partners are preferably selected from

- a) antibodies and fragments such as, for example, Fv, Fab, F(ab')₂, thereof
- b) protein-like compounds which interact, for example via the above leucine zipper region or another sequence section, with
 40 PARP, and
 - c) low molecular weight effectors which modulate a biological PARP function such as, for example, the catalytic PARP activity, i.e. NAD+-consuming ADP ribosylation, or the binding to an activator protein or to DNA.

45

The invention further relates to nucleic acids comprising

- a) a nucleotide sequence coding for at least one PARP homolog according to the invention, or the complementary nucleotide sequence thereof;
- b) a nucleotide sequence which hybridizes with a sequence as specified in a), preferably under stringent conditions; or
 - c) nucleotide sequences which are derived from the nucleotide sequences defined in a) and b) through the degeneracy of the genetic code.
- 10 Nucleic acids which are suitable according to the invention comprise in particular at least one of the partial sequences which code for the abovementioned amino acid sequence motifs.

Nucleic acids which are preferred according to the invention
15 comprise nucleotide sequences as shown in SEQ ID NO: 1 and 3,
and, in particular, partial sequences thereof which are
characteristic of PARP homologs according to the invention, such
as, for example, nucleotide sequences comprising

- 20 a) nucleotides +3 to +1715 shown in SEQ ID NO:1;
 - b) nucleotides +242 to +1843 shown in SEQ ID NO:3;
 - c) nucleotides +221 to +1843 shown in SEQ ID NO:5;
 - d) nucleotides +112 to +1710 shown in SEQ ID NO:7; or
 - e) nucleotides +1 to +1584 shown in SEQ ID NO:9

25

or partial sequences of a), b), c), d) and e) which code for the abovementioned characteristic amino acid sequence motifs of the PARP homologs according to the invention.

30 The invention further relates to expression cassettes which comprise at least one of the above-described nucleotide sequences according to the invention under the genetic control of regulatory nucleotide sequences. These can be used to prepare recombinant vectors according to the invention, such as, for 35 example, viral vectors or plasmids, which comprise at least one

35 example, viral vectors or plasmids, which comprise at least one expression cassette according to the invention.

Recombinant microorganisms according to the invention are transformed with at least one of the abovementioned vectors.

40

The invention also relates to transgenic mammals transfected with a vector according to the invention.

The invention further relates to an in vitro detection method, 45 which can be carried out homogeneously or heterogeneously, for PARP inhibitors, which comprises

- incubating an unsupported or supported poly-ADP-ribosylatable a) target with a reaction mixture comprising
 - al) a PARP homolog according to the invention;
 - a2) a PARP activator; and
- 5 a3) a PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected;
 - carrying out the polyADP ribosylation reaction; and b)
 - determining the polyADP ribosylation of the target qualitatively or quantitatively.

The detection method is preferably carried out by preincubating the PARP homolog with the PARP activator and the PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected, for example for about 1-30 minutes, before carrying out the poly-

15 ADP ribosylation reaction.

After activation by DNA with single strand breaks (referred to as "activated DNA" according to the invention), PARP poly-ADP ribosylates a large number of nuclear proteins in the presence of 20 NAD. These proteins include, on the one hand, PARP itself, but also histones etc.

The poly-ADP-ribosylatable target preferably used in the detection method is a histone protein in its native form or a poly-25 ADP-ribosylatable equivalent derived therefrom. A histone preparation supplied by Sigma (SIGMA, catalogue No. H-7755; histone type II-AS from calf thymus, Luck, J. M., et al., J. Biol. Chem., 233, 1407 (1958), Satake K., et al., J. Biol. Chem, 235, 2801 (1960)) was used by way of example. It is possible in principle 30 to use all types of proteins or parts thereof amenable to poly-ADP-ribosylation by PARP. These are preferably nuclear proteins, e.g. histones, DNA polymerase, telomerase or PARP itself. Synthetic peptides derived from the corresponding proteins can also act as target.

35

In the ELISA according to the invention it is possible to use amounts of histones in the range from about $0.1 \mu g/well$ to about 100 μg/well, preferably about 1 μg/well to about 10 μg/well. The amounts of the PARP enzyme are in a range from about 0.2 pmol/ 40 well to about 2 nmol/well, preferably from about 2 pmol/well to about 200 pmol/well, the reaction mixture comprising in each case 100 µg/well. Reductions to smaller wells and correspondingly

45 In the HTRF assay according to the invention, identical amounts of PARP are employed, and the amount of histone or modified histones is in the range from about 2 ng/well to about 25 µg/well,

smaller reaction volumes are possible.

preferably about 25 ng/well to about 2.5 μ g/well, the reaction mixture comprising in each case 50 μ l/well. Reductions to smaller wells and correspondingly smaller reaction volumes are possible.

5 The PARP activator used according to the invention is preferably activated DNA.

Various types of damaged DNA can function as activator. DNA damage can be produced by digestion with DNases or other DNA-modify10 ing enzymes (e.g. restriction endonucleases), by irradiation or other physical methods or chemical treatment of the DNA. It is further possible to simulate the DNA damage situation in a targeted manner using synthetic oligonucleotides. In the assays indicated by way of example, activated DNA from calf thymus was
15 employed (Sigma, product No. D4522; CAS: 91080-16-9, prepared by the method of Aposhian and Kornberg using calf thymus DNA (SIGMA D-1501) and deoxyribonuclease type I (D-4263). Aposhian H. V. and Kornberg A., J. Biol. Chem., 237, 519 (1962)). The activated DNA was used in a concentration range from 0.1 to 1000 μg/ml, prefera20 bly from 1 to 100 μg/ml, in the reaction step.

The polyADP ribosylation reaction is started in the method according to the invention by adding NAD $^+$. The NAD concentrations were in a range from about 0.1 μ M to about 10 mM, preferably in a 25 range from about 10 μ M to about 1 mM.

In the variant of the above method which can be carried out heterogeneously, the polyADP ribosylation of the supported target is determined using anti-poly(ADP-ribose) antibodies. To do this, 30 the reaction mixture is separated from the supported target, washed and incubated with the antibody. This antibody can itself be labeled. However, as an alternative for detecting bound anti-poly(ADP-ribose) antibody a labeled secondary antibody or a corresponding labeled antibody fragment may be applied. Suitable 35 labels are, for example, radiolabeling, chromophore- or fluoro-phore-labeling, biotinylation, chemiluminescence labeling, labeling with paramagnetic material or, in particular, enzyme labels, e.g. with horseradish peroxidase. Appropriate detection techniques are generally known to the skilled worker.

40

In the variant of the above process which can be carried out homogeneously, the unsupported target is labeled with an acceptor fluorophore. The target preferably used in this case is biotiny-lated histone, the acceptor fluorophore being coupled via avidin or streptavidin to the biotin groups of the histone. Particularly suitable as acceptor fluorophore are phycobiliproteins (e.g. phycocyanins, phycoerythrins), e.g. R-phycocyanin (R-PC), allophyco-

cyanin (APC), R-phycoerythrin (R-PE), C-phycocyanin (C-PC), B-phycoerythrin (B-PE) or their combinations with one another or with fluorescent dyes such as Cy5, Cy7 or Texas Red (Tandem system) (Thammapalerd, N. et al., Southeast Asian Journal of Tropical Medicine & Public Health, 27(2): 297-303 (1996); Kronick, M. N. et al., Clinical Chemistry, 29(9), 1582-1586 (1986); Hicks, J. M., Human Pathology, 15(2), 112-116 (1984)). The dye XL665 used in the examples is a crosslinked allophycocyanin (Glazer, A. N., Rev. Microbiol., 36, 173-198 (1982); Kronick, M. N., J. Imm.

10 Meth., 92, 1-13 (1986); MacColl, R. et al., Phycobiliproteins, CRC Press, Inc., Boca Raton, Florida (1987); MacColl, R. et al., Arch. Biochem. Biophys., 208(1), 42-48 (1981)).

It is additionally preferred in the homogeneous method to deter15 mine the polyADP ribosylation of the unsupported target using anti-poly(ADP-ribose) antibody which is labeled with a donor fluorophore which is able to transfer energy to the acceptor fluorophore when donor and acceptor are close in space owing to binding of the labeled antibody to the polyADP-ribosylated hi20 stone. A europium cryptate is preferably used as donor fluorophore for the anti-poly(ADP-ribose) antibody.

Besides the europium cryptate used, other compounds are also possible as potential donor molecules. This may entail, on the 25 one hand, modification of the cryptate cage. Replacement of the europium by other rare earth metals such as terbium is also conceivable. It is crucial that the fluorescence has a long duration to guarantee the time delay (Lopez, E. et al., Clin. Chem. 39/2, 196-201 (1993); US Patent 5,534,622).

30

The detection methods described above are based on the principle that there is a correlation between the PARP activity and the amount of ADP-ribose polymers formed on the histones. The assay described herein makes it possible to quantify the ADP-ribose

35 polymers using specific antibodies in the form of an ELISA and an HTRF (homogenous time-resolved fluorescence) assay. Specific embodiments of these two assays are described in detail in the following examples.

40 The developed HTRF (homogeneous time-resolved fluorescence) assay system measures the formation of poly(ADP-ribose) on histones using specific antibodies. In contrast to the ELISA, this assay is carried out in homogeneous phase without separation and washing steps. This makes a higher sample throughput and a smaller susceptibility to errors possible. HTRF is based on the fluor-escence resonance energy transfer (FRET) between two fluoro-

phores. In a FRET assay, an excited donor fluorophore can

transfer its energy to an acceptor fluorophore when the two are close to one another in space. In HTRF technology, the donor fluorophore is a europium cryptate [(Eu)K] and the acceptor is XL665, a stabilized allophycocyanin. The europium cryptate is based on studies by Jean Marie Lehn (Strasbourg) (Lopez, E. et al., Clin. Chem. 39/2, 196-201 (1993); US Patent 5,534,622).

In a homogeneous assay, all the components are also present during the measurement. Whereas this has advantages for carrying out the assay (rapidity, complexity), it is necessary to preclude interference by assay components (inherent fluorescence, quenching by dyes etc.). HTRF precludes such interference by time-delayed measurement at two wavelengths (665 nm, 620 nm). The HTRF has a very long decay time and time-delayed measurement is therefore possible. There is no longer any interference from short-lived background fluorescence (e.g. from assay components or inhibitors of the substance library). In addition, measurement is always carried out at two wavelengths in order to compensate for quench effects of colored substances. HTRF assays can be carried out, for example, in 96- or 384-well microtiter plate format and are evaluated using a discovery HTRF microplate analyzer (Canberra Packard).

Also provided according to the invention are the following in 25 vitro screening methods for binding partners for PARP, in particular for a PARP homolog according to the invention.

A first variant is carried out by

- al) immobilizing at least one PARP homolog on a support;
- 30 bl) contacting the immobilized PARP homolog with an analyte in which at least one binding partner is suspected; and
 - c1) determining, where appropriate after an incubation period, analyte constituents bound to the immobilized PARP homolog.

35 A second variant entails

- a2) immobilizing on a support an analyte which comprises at least one possible binding partner for the PARP homolog;
- b2) contacting the immobilized analyte with at least one PARP homolog for which a binding partner is sought; and
- 40 c3) examining the immobilized analyte, where appropriate after an incubation period, for binding of the PARP homolog.

The invention also relates to a method for the qualitative or quantitative determination of a nucleic acid encoding a PARP 45 homolog, which comprises

- a) incubating a biological sample with a defined amount of an exogenous nucleic acid according to the invention (e.g. with a length of about 20 to 500 bases or longer), hybridizing, preferably under stringent conditions, determining the hybridizing nucleic acids and, where appropriate, comparing with a standard; or
- b) incubating a biological sample with a defined amount of oligonucleotide primer pairs with specificity for a PARP
 10 homolog-encoding nucleic acid, amplifying the nucleic acid, determining the amplification product and, where appropriate, comparing with a standard.

The invention further relates to a method for the qualitative or 15 quantitative determination of a PARP homolog according to the invention, which comprises

- a) incubating a biological sample with at least one binding partner specific for a PARP homolog,
- b) detecting the binding partner/PARP complex and, whereappropriate,
 - c) comparing the result with a standard.

The binding partner in this case is preferably an anti-PARP antibody or a binding fragment thereof, which carries a 25 detectable label where appropriate.

The determination methods according to the invention for PARP, in particular for PARP homologs and for the coding nucleic acid sequences thereof, are suitable and advantageous for diagnosing 30 sepsis- or ischemia-related tissue damage, in particular strokes, myocardial infarcts, diabetes or septic shock.

The invention further comprises a method for determining the efficacy of PARP effectors, which comprises

- 35 a) incubating a PARP homolog according to the invention with an analyte which comprises an effector of a physiological or pathological PARP activity; removing the effector again where appropriate; and
- b) determining the activity of the PARP homolog, whereappropriate after adding substrates or cosubstrates.

The invention further relates to gene therapy compositions which comprise in a vehicle acceptable for gene therapy a nucleic acid construct which

45 a) comprises an antisense nucleic acid against a coding nucleic acid according to the invention; or

- b) a ribozyme against a noncoding nucleic acid according to the invention; or
- c) codes for a specific PARP inhibitor.
- 5 The invention further relates to pharmaceutical compositions comprising, in a pharmaceutically acceptable vehicle, at least one PARP protein according to the invention, at least one PARP binding partner according to the invention or at least one coding nucleotide sequence according to the invention.

- Finally, the invention relates to the use of binding partners of a PARP homolog for the diagnosis or therapy of pathological states in the development and/or progress of which at least one PARP protein, in particular a PARP homolog according to the invention, or a polypeptide derived therefrom, is involved. The
 - binding partner used can be, for example, a low molecular weight binding partner whose molecular weight can be, for example, less than about 2000 dalton or less than about 1000 dalton.
- 20 The invention additionally relates to the use of PARP binding partners for the diagnosis or therapy of pathological states mediated by an energy deficit. An energy deficit for the purpose of the present invention is, in particular, a cellular energy deficit which is to be observed in the unwell patient systemically or in individual body regions, organs or organ regions, or tissues or tissue regions. This is characterized by an NAD and/or ATP depletion going beyond (above or below) the physiological range of variation of the NAD and/or ATP level and mediated preferably by
- a protein with PARP activity, in particular a PARP homolog ac-30 cording to the invention, or a polypeptide derived therefrom.
- "Energy deficit-mediated disorders" for the purpose of the invention additionally comprise those in which tissue damage is attributable to cell death resulting from necrosis or apoptosis. The 35 methods according to the invention are suitable for treating and preventing tissue damage resulting from cell damage due to apo
 - preventing tissue damage resulting from cell damage due to apoptosis or necrosis; damage to nerve tissue due to ischemias and/or reperfusion; neurological disorders; neurodegenerative disorders; vascular stroke; for treating and preventing cardiovascular
- 40 disorders; for treating other disorders or conditions such as, for example, age-related macular degeneration, AIDS or other immunodeficiency disorders; arthritis; atherosclerosis; cachexia; cancer; degenerative disorders of the skeletal muscles; diabetes; cranial trauma; inflammatory disorders of the gastrointestinal
- 45 tract such as, for example, Crohn's disease; muscular dystrophy; osteoarthritis; osteoporosis; chronic and/or acute pain; kidney failure; retinal ischemia; septic shock (such as, for example,

endotoxin shock); aging of the skin or aging in general; general manifestations of aging. The methods according to the invention can additionally be employed for extending the life and the proliferative capacity of body cells and for sensitizing tumor cells in connection with irradiation therapy.

The invention particularly relates to the use of a PARP binding partner as defined above for the diagnosis or therapy (acute or prophylactic) of pathological states mediated by energy deficits 10 and selected from neurodegenerative disorders, or tissue damage caused by sepsis or ischemia, in particular of neurotoxic disturbances, strokes, myocardial infarcts, damage during or after infarct lysis (e.g. with TPA, Reteplase or mechanically with laser or Rotablator) and of microinfarcts during and after heart 15 valve replacement, aneurysm resections and heart transplants, trauma to the head and spinal cord, infarcts of the kidney (acute kidney failure, acute renal insufficiency or damage during and after kidney transplant), damages of skeletal muscle, infarcts of the liver (liver failure, damage during or after a liver trans-20 plant), peripheral neuropathies, AIDS dementia, septic shock, diabetes, neurodegenerative disorders occuring after ischemia, trauma (craniocerebral trauma), massive bleeding, subarachnoid hemorrhages and stroke, as well as neurodegenerative disorders like Alzheimer's disease, multi-infarct dementia, Huntington's 25 disease, Parkinson's disease, amyotrophic lateral sclerosis, epilepsy, especially of generalized epileptic seizures such as petit mal and tonoclonic seizures and partial epileptic seizures, such as temporal lobe, and complex partial seizures, kidney failure, also in the chemotherapy of tumors and prevention of meta-30 stasis and for the treatment of inflammations and rheumatic disorders, e.g. of rheumatoid arthritis; further for the treatment of revascularization of critically narrowed coronary arteries and critically narrowed peripheral arteries, e.g. leg arteries.

"Ischemia" comprises for the purposes of the invention a localized undersupply of oxygen to a tissue, caused by blockage of arterial blood flow. Global ischemia occurs when the blood flow to the entire brain is interrupted for a limited period. This may be caused, for example, by cardiac arrest. Focal ischemia occurs
When part of the brain is cut off from its normal blood supply. Focal ischemia may be caused by thromboembolic closure of a blood vessel, by cerebral trauma, edemas or brain tumor. Even transient ischemias can lead to wideranging neuronal damage. Although damage to "nerve tissue" may occur days or weeks after the start of the ischemia, some permanent damage (e.g. necrotic cell death) occurs in the first few minutes after interruption of the blood supply. This damage is caused, for example, by the neurotoxicity

of glutamate and follows secondary reperfusion, such as, for example, release of free radicals (e.g. oxygen free radicals, NO free radicals). Ischemias may likewise occur in other organs and tissues such as, for example, in the heart (myocardial infarct and other cardiovascular disorders caused by occlusion of the coronary arteries) or in the eye (ischemia of the retina).

The invention additionally relates to the use of an effective therapeutic amount of a PARP binding partner for influencing neu10 ronal activity. "Neuronal activity" for the purposes of the invention may consist of stimulation of damaged neurons, promotion of neuronal regeneration or treatment of neuronal disorders.

"Neuronal damage" for the purposes of the invention comprises

15 every type of damage to "nerve tissue" and every physical or mental impairment or death resulting from this damage. The cause of the damage may be, for example, metabolic, toxic, chemical or thermal in nature and includes by way of example ischemias, hypoxias, trauma, cerebrovascular damage, operations, pressure, hemorrhages, irradiation, vasospasms, neurodegenerative disorders, infections, epilepsy, perception disorders, disturbances of glutamate metabolism and the secondary effects caused thereby.

"Nerve tissue" for the purposes of the invention comprises the
25 various components forming the nervous system, consisting of, inter alia, neurons, glia cells, astrocytes, Schwann cells, the vascular system inside and for supplying, the CNS, brain, brain stem, spinal cord, peripheral nervous system etc.

- 30 "Neuroprotective" for the purposes of the invention comprises the reduction, the cessation, the slowing down or the improvement of neuronal damage and the protection, the restoration and the regeneration of nerve tissue which was exposed to neuronal damage.
- 35 "Prevention of neurodegenerative disorders" includes the possibility of preventing, slowing down and improving neurodegenerative disorders in people for whom such a disorder has been diagnosed or who are included in appropriate risk groups for these neurodegenerative disorders. Treatments for people already suffering 40 from symptoms of these disorders are likewise meant.

"Treatment" for the purposes of the invention comprises

(i) preventing a disorder, a disturbance or a condition in45 people with a predisposition thereto;

- (iii) improving a disorder, a disturbance or a condition.

Examples of "neurological disorders" which can be treated by the methods according to the invention are neuralgias (trigeminal, glossopharyngeal), myasthenia gravis, muscular dystrophies, amyorophic lateral sclerosis (ALS), progressive muscular atrophy, pe-10 ripheral neuropathies caused by poisoning (e.g. lead poisoning), Guillain-Barré syndrome, Huntington's disease, Alzheimer's disease, Parkinson's disease, or plexus disorders. The methods according to the invention are preferably suitable for treating neurological disorders selected from peripheral neuropathies cau-15 sed by physical injury or illness; cranial trauma such as, for example, traumatic brain injury; physical damage to the spinal cord; stroke associated with brain damage, such as vascular stroke in conjunction with hypoxia and brain damage, and cerebral reperfusion damage; demyelinating disorders (myelopathies, Alz-20 heimer's disease, Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis).

The methods according to the invention can additionally be used for treating cardiovascular disorders. "Cardiovascular disorders"

25 for the purposes of the invention comprise those which cause ischemias or are caused by ischemias or ischemia/reperfusion of the heart. Examples are coronary vessel disorders (for example atherosclerosis), angina pectoris, myocardial infarct, cardiovascular damage due to cardiac arrest or bypass operation.

30

5

The methods according to the invention can be used for treating cancer or for sensitizing cancer cells for irradiation therapy. The term "cancer" is to be understood in the widest sense. Modulators of the proteins according to the invention can be used as 35 "anti-cancer therapy agents". For example, the methods can be used for treating types of cancer or tumor cells, such as ACTHproducing tumors, acute lymphatic or lymphoblastic leukemia; acute or chronic lymphocytic leukemia; acute nonlymphocytic leukemia; bladder cancer; brain tumors; breast cancer; cervical car-40 cinoma; chronic myelocytic leukemia; bowel cancer; T-zone lymphoma; endometriosis; esophageal cancer; gall bladder cancer; Ewing's sarcoma; head and neck cancer; cancer of the tongue; Hodgkin's lymphoma; Kaposi's sarcoma; renal cancer; liver cancer; lung cancer; mesothelioma; multiple myeloma; neuroblastoma; non-45 Hodgkin lymphoma; osteosarcoma; ovarian carcinoma; glioblastoma; mammary carcinoma; cervical carcinoma; prostate cancer; pancreatic cancer; penis cancer; retinoblastoma; skin cancer; stomach

cancer; thyroid cancer; uterine carcinoma; vaginal carcinoma; Wilm's tumor; or trophoblastoma.

"Radiosensitizer" or "irradiation sensitizer" for the purposes of 5 the invention relates to molecules which increase the sensitivity of the cells in the body to irradiation with electromagnetic radiation (for example X-rays) or speed up this irradiation treatment. Irradiation sensitizers increase the sensitivity of cancer cells to the toxic effects of the electromagnetic radiation.

10 Those disclosed in the literature include mitomycin C, 5-bromodeoxyuridine and metronidazole. It is possible to use radiation with wavelengths in the range from 10^{-20} to 10 meters, preferably gamma rays (10^{-20} to 10^{-13} m), X-rays (10^{-11} to 10^{-9} m), ultraviolet radiation (10 nm to 400 nm), visible light (400 nm to 700 nm),

15 infrared radiation (700 nm to 1 mm) and microwave radiation (1 mm to 30 cm).

Disorders which can be treated by such a therapy are, in particular, neoplastic disorders, benign or malignant tumors and cancer. 20 The treatment of other disorders using electromagnetic radiation is likewise possible.

The present invention will now be described in more detail with reference to the appended figures. These show:

In Figure 1 a sequence alignment of human PARP (human PARP1) and two PARPs preferred according to the invention (human PARP2, human PARP3, murine PARP3). Sequence agreements between human PARP1 and human PARP2, human PARP3 or murine PARP3 are depicted within frames. The majority sequence is indicated over the alignment. The zinc finger motifs of human PARP1 are located in the sequence sections corresponding to amino acid residues 21 to 56 and 125 to 162;

35 In Figure 2 Northern blots with various human tissues to illustrate the tissue distribution of PARP2 and PARP3 molecules according to the invention. Lane 1: brain; lane 2: heart; lane 3: skeletal muscle; lane 4: colon; lane 5: thymus; lane 6: spleen; lane 7: kidney; lane 8: liver; lane 9: intestine; lane 10: pla-40 centa; lane 11: lung; lane 12: peripheral blood leukocytes; the respective position of the size standard (kb) is indicated.

In Figure 3 a Northern blot with further various human tissues to illustrate the tissue distribution of the PARP3 molecule accord45 ing to the invention. Lane 1: heart; lane 2: brain; lane 3: placenta; lane 4: lung; lane 5: liver; lane 6: skeletal muscle; lane

7: kidney; lane 8: pancreas; the respective position of the size standard (kb) is indicated.

In Figure 4 a Western blot with various human tissues to illus5 trate the tissue distribution of the PARP3 molecule according to
the invention at the protein level. Lane 1: heart; lane 2: lung;
lane 3: liver; lane 4: spleen; lane 5: kidney; lane 6: colon;
lane 7: muscle; lane 8: brain; the respective position of the
size standard (kD) is indicated.

10

In Figure 5 a Western blot with various human tissues to illustrate the tissue distribution of the PARP3 molecule according to the invention. Lane 1: frontal cortex; lane 2: posterior cortex; lane 3: cerebellum; lane 4: hippocampus; lane 5: olfactory bulb; lane 6: striatum; lane 7: thalamus; lane 8: midbrain; lane 9: entorhinal cortex; lane 10: pons; lane 11: medulla; lane 12: spinal cord.

In Figure 6 a diagrammatic representation of the PARP assay 20 (ELISA)

In Figure 7 a diagrammatic representation of the PARP assay (HTRF)

25 Further preferred embodiments of the invention are described in the following sections.

PARP homologs and functional equivalents

30 Unless stated otherwise, for the purposes of the present description amino acid sequences are indicated starting with the N terminus. If the one-letter code is used for amino acids, then G is glycine, A is alanine, V is valine, L is leucine, I is isoleucine, S is serine, T is threonine, D is aspartic acid, N is asparagine, E is glutamic acid, Q is glutamine, W is tryptophan, H is histidine, R is arginine, P is proline, K is lysine, Y is tyrosine, F is phenylalanine, C is cysteine and M is methionine.

The present invention is not confined to the PARP homologs
40 specifically described above. On the contrary, those homologs which are functional equivalents thereof are also embraced. Functional equivalents comprise both natural, such as, for example, species-specific or organ-specific, and artificially produced variants of the proteins specifically described herein.
45 Functional equivalents according to the invention differ by

addition, substitution, inversion, insertion and/or deletion of one or more amino acid residues of human PARP2 (SEQ ID NO:2),

19

human PARP3 (SEQ ID NO: 4 and 6) and mouse PARP3 (SEQ ID:8 and 10), there being at least retention of the NAD-binding function of the protein mediated by a functional catalytic C-terminal domain. Likewise, the poly(ADP-ribose)-producing catalytic 5 activity should preferably be retained. Functional equivalents also comprise where appropriate those variants in which the region similar to the leucine zipper is essentially retained.

It is moreover possible, for example, starting from the sequence for human PARP2 or human PARP3 to replace certain amino acids by those with similar physicochemical properties (bulk, basicity, hydrophobicity, etc.). It is possible, for example, for arginine residues to be replaced by lysine residues, valine residues by isoleucine residues or aspartic acid residues by glutamic acid residues. However, it is also possible for one or more amino acids to be exchanged in sequence, added or deleted, or several of these measures can be combined together. The proteins which have been modified in this way from the human PARP2 or human PARP3 sequence have at least 60%, preferably at least 75%, very particularly preferably at least 85%, homology with the starting sequence, calculated using the algorithm of Pearson and Lipman, Proc. Natl. Acad. Sci (USA) 85(8), 1988, 2444-2448.

The following homologies have been determined at the amino acid 25 level and DNA level between human PARP1, 2 and 3 (FastA program, Pearson and Lipman, loc. cit.):

Amino acid homologies:

20			
30		Percent identity	Percent identity
			in
			PARP signature
35	PARP1/PARP2	41.97% (517)	86% (50)
	PARP1/PARP3	33.81% (565)	53.1% (49)
	PARP2/PARP3	35.20% (537)	53.1% (49)

40 Numbers in parentheses indicate the number of overlapping amino acids.

DNA Homologies:

5		Percent identity in the ORF	Percent identity in PARP signature				
	PARP1/PARP2	60.81% (467)	77.85% (149)				
10	PARP1/PARP3	58.81% (420)	59.02% (61)				
.10	PARP2/PARP3	60.22% (269)	86.36% (22)				

Numbers in parentheses indicate the number of overlapping nucleotides.

15

The polypeptides according to the invention can be classified as homologous poly(ADP-ribose) polymerases on the basis of the great similarity in the region of the catalytic domain.

 20 It is also essential to the invention that the novel PARP homologs do not have conventional zinc finger motifs. This means that these enzymes are not necessarily involved in DNA repair or are so in a way which differs from PARP1, but are still able to carry out their pathological mechanism (NAD+ consumption and thus 25 energy consumption due to ATP consumption). The strong protein expression, particularly of PARP3, observable in the Western blot suggests a significant role in the NAD consumption. This is particularly important for drug development. Potential novel inhibitors of the polymerases according to the invention can thus 30 inhibit the pathological functions without having adverse effects on the desired physiological properties. This was impossible with inhibitors against the PARPs known to date since there was always also inhibition of the DNA repair function. The potentially mutagenic effect of known PARP inhibitors is thus easy to 35 understand. It is also conceivable to design PARP inhibitors so that they efficiently inhibit all PARP homologs with high affinity. In this case, a potentiated effect is conceivable where appropriate.

The PARP homolog which is preferred according to the invention and is shown in SEQ ID NO:2 (human PARP2) can advantageously be isolated from human brain, heart, skeletal muscle, kidney and liver. The expression of human PARP2 in other tissues or organs is distinctly weaker.

45

The PARP homolog which is preferred according to the invention and is shown in SEQ ID NO: 4 and 6 (human PARP3) can advantageously be isolated from human brain (in this case very preferentially from the hippocampus), heart, skeletal muscle, 5 liver or kidney. The expression of human PARP3 in other tissues or organs, such as muscle or liver, is distinctly weaker.

The skilled worker familiar with protein isolation will make use of the combination of preparative methodologies which is most suitable in each case for isolating natural PARPs according to the invention from tissues or recombinantly prepared PARPs according to the invention from cell cultures. Suitable standard preparative methods are described, for example, in Cooper, T.G., Biochemische Arbeitsmethoden, published by Walter de Gruyter, 15 Berlin, New York or in Scopes, R. Protein Purification, Springer Verlag, New York, Heidelberg, Berlin.

The invention additionally relates to PARP2 and PARP3 homologs which, although they can be isolated from other eukaryotic

20 species, i.e. invertebrates or vertebrates, especially other mammals such as, for example, mice, rats, cats, dogs, pigs, sheep, cattle, horses or monkeys, or from other organs such as, for example the myocardium, have the essential structural and functional properties predetermined by the PARPs according to the invention.

In particular, the human PARP2 which can be isolated from human brain, and its functional equivalents, are preferred agents for developing inhibitors of neurodegenerative diseases as for 30 example stroke. This is because it can be assumed that drug development based on PARP2 as indicator makes it possible to develop inhibitors which are optimized for use in the human brain. However, it cannot be ruled out that inhibitors developed on the basis of PARP2 can also be employed for treating 35 PARP-mediated pathological states in other organs, too (see tissue distribution of the proteins according to the invention).

PARP2 and presumably PARP3 are also, similar to PARP1, activated by damaged DNA, although by a presumably different mechanism.

40 Significance in DNA repair is conceivable. Blockade of the PARPs according to the invention would also be beneficial in indications such as cancer (e.g. in the radiosensitization of tumor patients).

45 Another essential biological property of PARPs according to the invention and their functional equivalents is to be seen in their ability to bind an interacting partner. Human PARP2 and 3 differ

from previously disclosed PARPs from higher eukaryotes such as, in particular, mammals by having potential so-called leucine zipper motifs. This is a typical motif for protein-protein interactions. It is possible that these motifs permit modulation of PARP activity by an interacting partner. This additional structural element thus also provides a possible starting point for development of PARP effectors such as, for example, inhibitors.

10 The invention thus further relates to proteins which interact with PARP2 and/or 3, preferably those which bring about their activation or inactivation.

The invention further relates to proteins which still have the 15 abovementioned ligand-binding activity and which can be prepared starting from the specifically disclosed amino acid sequences by targeted modifications.

It is possible, starting from the peptide sequence of the
proteins according to the invention, to generate synthetic
peptides which are employed, singly or in combination, as
antigens for producing polyclonal or monoclonal antibodies. It is
also possible to employ the PARP protein or fragments thereof for
generating antibodies. The invention thus also relates to peptide
fragments of PARP proteins according to the invention which
comprise characteristic partial sequences, in particular those
oligo- or polypeptides which comprise at least one of the
abovementioned sequence motifs. Fragments of this type can be
obtained, for example, by proteolytic digestion of PARP proteins
or by chemical synthesis of peptides.

Novel specific PARP2 and PARP3 binding partners

Active and preferably selective inhibitors against the proteins 35 according to the invention were developed using the specific assay systems described above for binding partners for PARP2 and PARP3. These inhibitors optionally are also active vis a vis PARP1.

- 40 Inhibitors provided according to the invention have a strong inhibitory activity on PARP2. The $\rm K_i$ values may in this case be less than about 1000 nM, such as less than about 700 nM, less than about 200 nM or less than about 30 nM, e.g. about 1 to 20 nM.
- 45 Inhibitors according to the invention may also have a surprising selectivity for PARP2. This is shown by the $K_i(PARP1)$: $K_i(PARP2)$ ratio for such inhibitors according to the invention which is,

for example, greater than 3 or greater than 5, as for example greater than 10 or greater than 20.

An example which should be mentioned is 4-(N-(4-hydroxyphe-5 nyl)aminomethyl)-(2H)-dihydrophthalazine-1-one. The preparation of this and other analogous compounds may be performed according to Puodzhyunas et al., Pharm. Chem. J. 1973, 7, 566 or Mazkanowa et al., Zh. Obshch. Khim., 1958, 28, 2798, or Mohamed et al., Ind. J. Chem. B., 1994, 33, 769 each incorporated by reference.

10

The above identified compuound shows a Ki value of 113 nM for PARP2 and is eight times more selective for PARP2 than for PARP3.

Nucleic acids coding for PARP homologs:

15

Unless stated otherwise, nucleotide sequences are indicated in the present description from the 5' to the 3' direction.

The invention further relates to nucleic acid sequences which

20 code for the abovementioned proteins, in particular for those
having the amino acid sequence depicted in SEQ ID NO: 2, 4, 6, 8
and 10, but without being restricted thereto. Nucleic acid
sequences which can be used according to the invention also
comprise allelic variants which, as described above for the amino

25 acid sequences, are obtainable by deletion, inversion, insertion,
addition and/or substitution of nucleotides, preferably of
nucleotides shown in SEQ ID NO: 1, 3, 7 and 9, but with essential
retention of the biological properties and the biological
activity of the corresponding gene product. Nucleotide sequences

30 which can be used are obtained, for example, by nucleotide
substitutions causing silent (without alteration of the amino
acid sequence) or conservative amino acid changes (exchange of
amino acids of the same size, charge, polarity or solubility).

35 Nucleic acid sequences according to the invention also embrace functional equivalents of the genes, such as eukaryotic homologs for example from invertebrates such as Caenorhabditis or Drosophila, or vertebrates, preferably from the mammals described above. Preferred genes are those from vertebrates which code for 40 a gene product which has the properties essential to the invention as described above.

The nucleic acids according to the invention can be obtained in a conventional way by various routes:

For example, a genomic or a cDNA library can be screened for DNA which codes for a PARP molecule or a part thereof. For example, a cDNA library obtained from human brain, heart or kidney can be screened with a suitable probe such as, for example, a labeled 5 single-stranded DNA fragment which corresponds to a partial sequence of suitable length selected from SEQ ID NO: 1 or 3, or sequence complementary thereto. For this purpose, it is possible, for example, for the DNA fragments of the library which have been transferred into a suitable cloning vector to be, after 10 transformation into a bacterium, plated out on agar plates. The clones can then be transferred to nitrocellulose filters and, after denaturation of the DNA, hybridized with the labeled probe. Positive clones are then isolated and characterized.

15 The DNA coding for PARP homologs according to the invention or partial fragments can also be synthesized chemically starting from the sequence information contained in the present application. For example, it is possible for this purpose for oligonucleotides with a length of about 100 bases to be
20 synthesized and sequentially ligated in a manner known per se by, for example, providing suitable terminal restriction cleavage sites.

The nucleotide sequences according to the invention can also be
25 prepared with the aid of the polymerase chain reaction (PCR). For
this, a target DNA such as, for example, DNA from a suitable
full-length clone is hybridized with a pair of synthetic
oligonucleotide primers which have a length of about 15 bases and
which bind to opposite ends of the target DNA. The sequence
30 section lying between them is then filled in with DNA polymerase.
Repetition of this cycle many times allows the target DNA to be
amplified (cf. White et al.(1989), Trends Genet. 5, 185).

The nucleic acid sequences according to the invention are also to 35 be understood to include truncated sequences, single-stranded DNA or RNA of the coding and noncoding, complementary DNA sequence, mRNA sequences and cDNAs derived therefrom.

The invention further embraces nucleotide sequences hybridizing
40 with the above sequences under stringent conditions. Stringent
hybridization conditions for the purpose of the present invention
exist when the hybridizing sequences have a homology of about 70
to 100%, such as, for example about 80 to 100% or 90 to 100%
(preferably in an amino acid section of at least about 40, such
45 as, for example, about 50, 100, 150, 200, 400 or 500 amino
acids).

Stringent conditions for the screening of DNA, in particular cDNA banks, exist, for example, when the hybridization mixture is washed with 0.1X SSC buffer (20X SSC buffer = 3M NaCl, 0.3M sodium citrate, pH 7.0) and 0.1% SDS at a temperature of about 5 60°C.

Northern blot analyses are analyses are washed under stringent conditions with 0.1% SSC, 0,1% SDS at a temperature of about 65°C, for example.

10

Nucleic acid derivatives and expression constructs:

The nucleic acid sequences are also to be understood to include derivatives such as, for example, promoter variants or

15 alternative splicing variants. The promoters operatively linked upstream of the nucleotide sequences according to the invention may moreover be modified by nucleotide addition(s) or substitution(s), inversion(s), insertion(s) and/or deletion(s), but without impairing the functionality or activity of the

20 promoters. The promoters can also have their activity increased by modifying their sequence, or be completely replaced by more effective promoters even from heterologous organisms. The promoter variants described above are used to prepare expression cassettes according to the invention.

25

Specific examples of human PARP2 splicing variants which may be mentioned are:

Variant human PARP2a: Deletion of base pairs 766 to 904 (cf. SEQ 30 ID NO:1). This leads to a frame shift with a new stop codon ("TAA" corresponding to nucleotides 922 to 924 in SEQ ID NO:1). Variant human PARP2b: Insertion of 5'- gta tgc cag gaa ggt cat ggg cca gca aaa ggg tct ctg -3' after nucleotide 204 (SEQ ID NO:1). This extends the amino acid 35 sequence by the insertion: GMPGRSWASKRVS

Nucleic acid derivatives also mean variants whose nucleotide sequences in the region from -1 to -1000 in front of the start codon have been modified so that gene expression and/or protein 40 expression is increased.

Besides the nucleotide sequence described above, the nucleic acid constructs which can be used according to the invention comprise in functional, operative linkage one or more other regulatory

45 sequences, such as promoters, amplification signals, enhancers, polyadenylation sequences, origins of replication, reporter genes, selectable marker genes and the like. This linkage may,

depending on the desired use, lead to an increase or decrease in gene expression.

In addition to the novel regulatory sequences, it is possible for 5 the natural regulatory sequence still to be present in front of the actual structural genes. This natural regulation can, where appropriate, be switched off by genetic modification, and the expression of the genes increased or decreased. However, the gene construct may also have a simpler structure, that is to say no 10 additional regulatory signals are inserted in front of the structural genes, and the natural promoter with its regulation is not deleted. Instead, the natural regulatory sequence is mutated in such a way that regulation no longer takes place, and gene expression is enhanced or diminished. It is also possible to 15 insert additional advantageous regulatory elements at the 3' end of the nucleic acid sequences. The nucleic acid sequences can be present in one or more copies in the gene construct.

Advantageous regulatory sequences for the expression method

20 according to the invention are, for example, present in promoters such as cos, tac, trp, tet, trp-tet, lpp, lac, lpp-lac, lacIq, T7, T5, T3, gal, trc, ara, SP6, l-PR or the l-PL promoter, which are advantageously used in Gram-negative bacteria. Other advantageous regulatory sequences are present, for example, in

25 the Gram-positive promoters amy and SPO2, in the yeast promoters ADC1, MFa, AC, P-60, CYC1, GAPDH or in the plant promoters CaMV/35S, SSU, OCS, lib4, usp, STLS1, B33, nos or in the ubiquitin or phaseolin promoter.

30 It is possible in principle to use all natural promoters with their regulatory sequences. It is also possible and advantageous to use synthetic promoters.

Said regulatory sequences are intended to make specific

35 expression of the nucleic acid sequences and protein expression possible. This may mean, for example, depending on the host organism that the gene is expressed or overexpressed only after induction, or that it is immediately expressed and/or overexpressed.

The regulatory sequences or factors may moreover preferably have a positive influence on, and thus increase or decrease, the expression. Thus, enhancement of the regulatory elements may advantageously take place at the level of transcription by using 45 strong transcription signals such as promoters and/or enhancers.

27

However, it is also possible to enhance translation by, for example, improving the stability of the mRNA.

Enhancers mean, for example, DNA sequences which bring about 5 increased expression via an improved interaction between RNA polymerase and DNA.

The recombinant nucleic acid construct or gene construct is, for expression in a suitable host organism, advantageously inserted

10 into a host-specific vector which makes optimal expression of the genes in the host possible. Vectors are well known to the skilled worker and are to be found, for example, in "Cloning Vectors" (Pouwels P. H. et al., Ed., Elsevier, Amsterdam-New York-Oxford, 1985). Apart from plasmids, vectors also mean all other vectors

15 known to the skilled worker, such as, for example, phages, viruses, such as SV40, CMV, baculovirus and adenovirus, transposons, IS elements, phasmids, cosmids, and linear or circular DNA. These vectors may undergo autonomous replication in the host organism or chromosomal replication.

20

Expression of the constructs:

The recombinant constructs according to the invention described above are advantageously introduced into a suitable host system 25 and are expressed. Cloning and transfection methods familiar to the skilled worker are preferably used in order to bring about expression of said nucleic acids in the particular expression system. Suitable systems are described, for example, in Current Protocols in Molecular Biology, F. Ausubel et al., ed., Wiley 30 Interscience, New York 1997.

Suitable host organisms are in principle all organisms which make it possible to express the nucleic acids according to the invention, their allelic variants, their functional equivalents or derivatives or the recombinant nucleic acid construct. Host organisms mean, for example, bacteria, fungi, yeasts, plant or animal cells. Preferred organisms are bacteria such as those of the genera Escherichia, such as, for example, Escherichia coli, Streptomyces, Bacillus or Pseudomonas, eukaryotic microorganisms such as Saccharomyces cerevisiae, Aspergillus, higher eukaryotic cells from animals or plants, for example Sf9 or CHO cells.

The gene product can also, if required, be expressed in transgenic organisms such as transgenic animals such as, in 45 particular, mice, sheep, or transgenic plants. The transgenic organisms may also be so-called knock-out animals or plants in which the corresponding endogenous gene has been switched off,

such as, for example, by mutation or partial or complete deletion.

The combination of the host organisms and the vectors appropriate ${\bf 5}$ for the organisms, such as plasmids, viruses or phages, such as, for example, plasmids with the RNA polymerase/promoter system, phages λ , μ or other temperate phages or transposons and/or other advantageous regulatory sequences forms an expression system. The term expression systems preferably means, for example, a 10 combination of mammalian cells such as CHO cells, and vectors, such as pcDNA3neo vector, which are suitable for mammalian cells.

As described above, the gene product can also be expressed advantageously in transgenic animals, e.g. mice, sheep, or 15 transgenic plants. It is likewise possible to program cell-free translation systems with the RNA derived from the nucleic acid.

The gene product can also be expressed in the form of therapeutically or diagnostically suitable fragments. To isolate 20 the recombinant protein it is possible and advantageous to use vector systems or oligonucleotides which extend the cDNA by certain nucleotide sequences and thus code for modified polypeptides which serve to simplify purification. Suitable modifications of this type are, for example, so-called tags which 25 act as anchors, such as, for example, the modification known as the hexa-histidine anchor, or epitopes which can be recognized as antigens by antibodies (described, for example, in Harlow, E. and Lane, D., 1988, Antibodies: A Laboratory Manual. Cold Spring Harbor (N.Y.) Press). These anchors can be used to attach the 30 proteins to a solid support such as, for example, a polymer matrix, which can, for example, be packed into a chromatography column, or to a microtiter plate or to another support.

These anchors can also at the same time be used to recognize the 35 proteins. It is also possible to use for recognition of the proteins conventional markers such as fluorescent dyes, enzyme markers which form a detectable reaction product after reaction with a substrate, or radioactive markers, alone or in combination with the anchors for derivatizing the proteins.

Production of antibodies:

40

Anti-PARP2 antibodies are produced in a manner familiar to the skilled worker. Antibodies mean both polyclonal, monoclonal, 45 human or humanized antibodies or fragments thereof, single chain antibodies or also synthetic antibodies, likewise antibody fragments such as Fv, Fab and F(ab')₂. Suitable production methods

are described, for example, in Campbell, A.M., Monoclonal Anti-body Technology, (1987) Elsevier Verlag, Amsterdam, New York, Oxford and in Breitling, F. and Dübel, S., Rekombinante Antikörper (1997), Spektrum Akademischer Verlag, Heidelberg.

5

Further use of the coding sequence:

The present cDNA additionally provides the basis for cloning the genomic sequence of the novel PARP genes. This also includes the 10 relevant regulatory or promoter sequence, which is available, for example, by sequencing the region located 5' upstream of the cDNA according to the invention or located in the introns of the genes. The cDNA sequence information is also the basis for producing antisense molecules or ribozymes with the aid of known 15 methods (cf. Jones, J.T. and Sallenger, B.A. (1997) Nat. Biotechnol. 15, 902; Nellen, W. and Lichtenstein, C. (1993) TIBS, 18, 419). The genomic DNA can likewise be used to produce the gene constructs described above.

20 Another possibility of using the nucleotide sequence or parts thereof is to generate transgenic animals. Transgenic overexpression or genetic knock-out of the sequence information in suitable animal models may provide further valuable information about the (patho)physiology of the novel genes.

25

Therapeutic applications:

In situations where there is a prevailing deficiency of a protein according to the invention it is possible to employ several

30 methods for replacement. On the one hand, the protein, natural or recombinant, can be administered directly or by gene therapy in the form of its coding nucleic acid (DNA or RNA). It is possible to use any suitable vectors for this, for example both viral and non-viral vehicles. Suitable methods are described, for example,

35 by Strauss and Barranger in Concepts in Gene Therapy (1997), Walter de Gruyter, publisher. Another alternative is provided by stimulation of the endogenous gene by suitable agents.

It is also possible to block the turnover or the inactivation of 40 PARPs according to the invention, for example by proteases. Finally, inhibitors or agonists of PARPs according to the invention can be employed.

In situations where a PARP is present in excess or is 45 overactivated, various types of inhibitors can be employed. This inhibition can be achieved both by antisense molecules, ribozymes, oligonucleotides or antibodies, and by low molecular weight compounds.

The active substances according to the invention, i.e. PARP pro-5 teins, nucleic acids and PARP binding partners such as, for example, antibodies or modulators, can be administered either as sinqle therapeutic active substances or as mixtures with other therapeutic active substances. They can be administered as such, but in general they are administered in the form of pharmaceutical 10 compositions, i.e. as mixtures of the active substance(s) with at least one suitable pharmaceutical carrier or diluent. The active substances or compositions can be administered in any way suitable for the particular therapeutic purpose, e.g. orally or parenterally.

15

The nature of the pharmaceutical composition and of the pharmaceutical carrier or diluent depends on the required mode of administration. Oral compositions can be, for example, in the form of tablets or capsules and may contain customary excipients such as 20 binders (e.g. sirup, acacia, gelatin, sorbitol, tragacanth or polyvinylpyrrolidone), bulking agents (e.g. lactose, sugar, corn starch, calcium phosphate, sorbitol or glycine), lubricants (e.g. magnesium stearate, talc, polyethylene glycol or silica), disintegrants (e.g. starch) or wetting agents (e.g. sodium lauryl sul-25 fate). Oral liquid products may be in the form of aqueous or oily suspensions, solutions, emulsions, sirups, elixirs or sprays etc. or may be in the form of dry powders for reconstitution with water or another suitable carrier. Liquid products of these types may contain conventional additives, for example suspending 30 agents, flavorings, diluents or emulsifiers. It is possible to employ for parenteral administration solutions or suspensions with conventional pharmaceutical carriers. Parenteral administration of active substances according to the invention advantageously takes place using a liquid pharmaceutical composition 35 which can be administered parenterally, in particular intravenously. This preferably contains an effective amount of at least one active substance, preferably in dissolved form, in a pharmaceutically acceptable carrier suitable for this purpose. Examples of pharmaceutical carriers suitable for this purpose are, in par-40 ticular, aqueous solutions such as, for example, physiological saline, phosphate-buffered saline, Ringer's solution, Ringer's lactate solution and the like. The composition may moreover contain further additions such as antioxidants, chelating agents or antimicrobial agents.

The choice in each case of the dosage of the active substances according to the invention and the particular dosage schedule are subject to a decision of the treating physician. The latter will select a suitable dose and an appropriate dosage schedule depending on the chosen route of administration, on the efficacy of the medicine in each case, on the nature and severity of the disorder to be treated, and on the condition of the patient and his response to the therapy. Thus, for example, the pharmacologically active substances can be administered to a mammal (human or animal) in doses of about 0.5 mg to about 100 mg per kg of body weight and day. They can be administered in a single dose or in several doses.

Nontherapeutic applications:

15

The nucleic acids according to the invention, such as, for example, cDNA, the genomic DNA, the promoter, and the polypeptide, and partial fragments thereof, can also be used in recombinant or nonrecombinant form for developing various test 20 systems.

For example, it is possible to establish a test system which is suitable for measuring the activity of the promoter or of the protein in the presence of a test substance. The methods of 25 measurement in this case are preferably simple ones, e.g. colorimetric, luminometric, fluorimetric, immunological or radioactive, and allow preferably a large number of test substances to be measured rapidly. Tests of this type are suitable and advantageous for so-called high-throughput screening. These test systems allow test substances to be assessed for their binding to or their agonism, antagonism or inhibition of proteins according to the invention.

Determination of the amount, activity and distribution of the 35 proteins according to the invention or their underlying mRNA in the human body can be used for the diagnosis, for the determination of the predisposition and for the monitoring of certain diseases. Likewise, the sequence of the cDNA and the genomic sequence may provide information about genetic causes of 40 and predispositions to certain diseases. It is possible to use for this purpose both DNA/RNA probes and antibodies of a wide variety of types. The nucleotide sequences according to the invention or parts thereof can further be used in the form of suitable probes for detecting point mutations, deletions or insertions.

The proteins according to the invention can further be used to identify and isolate their natural ligands or interacting partners. The proteins according to the invention can additionally be used to identify and isolate artificial or 5 synthetic liqands. For this purpose, the recombinantly prepared or purified natural protein can be derivatized in such a way that it has modifications which permit linkage to support materials. Proteins bound in this way can be incubated with various analytes, such as, for example, protein extracts or peptide 10 libraries or other sources of ligands. Specifically bound peptides, proteins or low molecular weight, non-proteinogenous substances can be isolated and characterized in this way. Non-proteinogenous substances mean, for example, low molecular weight chemical substances which may originate, for example, from 15 classical drug synthesis or from so-called substance libraries which have been synthesized combinatorially.

The protein extracts used are derived, for example, from homogenates of plants or parts of plants, microorganisms, human 20 or animal tissues or organs.

Ligands or interacting partners can also be identified by methods like the yeast two-hybrid system (Fields, S. and Song, O. (1989) Nature, 340, 245). The expression banks which can be employed in 25 this case may be derived, for example, from human tissues such as, for example, brain, heart, kidney etc.

The nucleic acid sequences according to the invention and the proteins encoded by them can be employed for developing reagents, 30 agonists and antagonists or inhibitors for the diagnosis and therapy of chronic and acute diseases associated with the expression or activation of one of the protein sequences according to the invention, such as, for example, with increased or decreased expression thereof. The reagents, agonists, 35 antagonists or inhibitors developed can subsequently be used to produce pharmaceutical preparations for the treatment or diagnosis of disorders. Examples of possible diseases in this connection are those of the brain, of the peripheral nervous system, of the cardiovascular system or of the eye, of septic 40 shock, of rheumatoid arthritis, diabetes, acute kidney failure, or of cancer.

The relevance of the proteins according to the invention for said indications was verified using specific inhibitors in relevant 45 animal models.

The invention is now illustrated in detail with reference to the following examples.

Example 1: Isolation of the PARP2 and PARP3 cDNA

The present cDNA sequences were found for the first time on sequence analysis of cDNA clones of a cDNA library from human brain (Human Brain 5'Stretch Plus cDNA Library, # HL3002a, Clontech). The mouse PARP3 clones were isolated from a "lambda triplex mouse brain cDNA library" (Clontech order No. ML5004t).

The sequences of these clones are described in SEQ ID NO:1, 3, 7 and 9.

Example 2: Expression of PARP2 and PARP3 in human tissues

The expression of human PARP2 and human PARP3 was investigated in twelve different human tissues by Northern blot analysis. A Human Multiple Tissue Northern Blot (MTN™) supplied by Clontech (#7760-1 and #7780-1) was hybridized for this purpose with an RNA probe. The probe was produced by in vitro transcription of the corresponding cDNA of human PARP2 and human PARP3 in the presence of digoxigenin-labeled nucleotides in accordance with the manufacturer's method (BOEHRINGER MANNHEIM DIG Easy Hyb order No. 1603 558, DIG Easy Hyb method for RNA:RNA hybridization). The protocol was modified to carry out the prehybridization: 2x1h with addition of herring sperm DNA (10 mg/ml of hybridization solution). Hybridization then took place overnight with addition of herring sperm DNA (10 mg/ml of hybridization solution). The bands were detected using the CDP-Star protocol (BOEHRINGER

After stringent washing, the transcript of PARP2 was mainly detected in human brain, heart, skeletal muscle, kidney and liver. The transcript size of about 1.9 kb corresponds to the 35 length of the cDNA determined (1.85kb) (cf. Figure 2(A)).

In other tissues or organs, human PARP2 expression is considerably weaker.

30 MANNHEIM CDP-Star™ order No. 1685 627).

40 After stringent washing, the transcript of PARP3 was mainly detected in heart, brain, kidney, skeletal muscle and liver. Expression in other tissues (placenta, lung, pancreas) is distinctly weaker (cf. Figure 2(B)). There are at least 2 transcripts for human PARP3, which can presumably be explained by different polyadenylation sites or alternative splicing. Their size (about 2.2 kb and 2.5 kb respectively) corresponds to the length of the cDNA determined (2.3kb). Washing was carried out

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with 0.2 x SSC/0.2% SDS at room temperature for 2 x 15 minutes and then with 0.1 x SSC/0.1% SDS at 65°C for 2 x 15 minutes (prepared from 20% SSC: 3M NaCl, 0.3M sodium citrate, pH 7.0).

5 Example 3: Production of antibodies

Specific antibodies against the proteins according to the invention were produced. These were used inter alia for analyzing the tissue distribution at the protein level of PARP2 and PARP3 by

10 immunoblot (Western blot) analysis. Examples of the production of such antibodies are indicated below.

The following peptides were prepared by synthesis in the manner familiar to the skilled worker for the antibody production. In 15 some cases, a cysteine residue was attached to the N or C terminals of the sequences in order to facilitate coupling to KLH (keyhole limpet hemocyanin).

PARP-2: NH₂-MAARRRSTGGGRARALNES-CO₂H (amino acids 1-20;

SEQ ID NO: 23)

NH₂-KTELQSPEHPLDQHYRNLHC-CO₂H (amino acids 335-353;

SEQ ID NO: 24)

PARP-3: NH₂-CKGRQAGREEDPFRSTAEALK-CO₂H (amino acids 25-44 SEQ ID NO: 25)
NH₂-CKQQIARGFEALEALEEALK-CO₂H (amino acids 230-248; SEQ ID NO: 26)

The production of an anti-PARP3 antibody is described as a representative example.

For human PARP3, polyclonal antibodies were raised in rabbits using a synthetic peptide having the peptide sequence H₂N-KQQIARG-FEALEALEEALK-CO₂H (SEQ ID NO: 27) (amino acids 230-248 of the human PARP3 protein sequence). The corresponding mouse sequence differs in this region only by one amino acid (H₂N-KQQIARGFEALEALEEAMK-CO₂H; SEQ ID NO: 28). A cysteine was also attached to the N terminus in order to make it possible for the protein to couple to KLH.

40 Rabbits were immunized a total of five times, at intervals of 7-14 days, with the KLH-peptide conjugate. The antiserum obtained was affinity-purified using the antigen. The specific IgG fraction was isolated from the serum using the respective peptides which, for this purpose, were initially immobilized on an affinity column in the manner familiar to the skilled worker. The respective antiserum was loaded onto this affinity column, and non-

specifically sorbed proteins were eluted with buffer. The spe-

cifically bound IgG fraction was eluted with 0.2 M glycine/HCl buffer pH 2.2. The pH was immediately increased using a 1M TRIS/ HCl buffer pH 7.5. The eluate containing the IgG fraction was mixed 1:1 (volume) with saturated ammonium sulfate solution and 5 incubated at $+4^{\circ}$ C for 30 min to complete the precipitation. The resulting precipitate was centrifuged at 10,000 g and, after removal of the supernatant, dissolved in the minimum amount of PBS/ TBS. The resulting solution was then dialyzed against PBS/TBS in the ratio 1:100 (volume). The antibodies were adjusted to a concentration of about 100 μ g of IgG/ml. The PARP3 antibodies purified in this way had high specificity for PARP3. Whereas mouse PARP3 was recognized well, there was no observable cross-reaction with PARP1 or PARP2.

15 Example 4: Analysis of the tissue distribution by immunoblot (Western blot)

The tissue distribution at the protein level was also investigated for PARP2 and PARP3 by immunoblot (Western blot) analysis.

Preparation of the mouse tissues for protein gels:

Tissues or cells were homogenized using a Potter or Ultra-Turrax. For this, 0.5 g of tissue (or cells) was incubated in 5 ml of

25 buffer (10 mM Tris-HCl pH 7.5, 1 mM EDTA, 6 mM MgCl₂), one tablet of protease inhibitor cocktail (Boehringer Mannheim, order No.: 1836153) and benzonase (purity grade I, MERCK) at 37°C for 30 min. Tissue samples from mice were produced for heart, lung, liver, spleen, kidney, intestine, muscle, brain and for human embryonic

30 kidney cells (HEK293, human embryonal kidney).

Protein gels:

20

The NuPAGE system supplied by NOVEX was used according to the instructions for protein gels. Polyacrylamide gels (NuPAGE 4-12% BisTris, NOVEX NP 0321), running buffer (MES-Running Buffer, NOVEX NP 0002), antioxidant (NOVEX NP 0005), protein size standard (Multi Mark Multi Colored Standard, NOVEX LC 5725), sample buffer (NuPAGE LDS Sample Buffer (4X), NOVEX NP 0007) were used.

Western blot:

Western blots were carried out using the NOVEX system in accordance with instructions. A nitrocellulose membrane (Nitrocellulose Pore size 45 μ m, NOVEX LC 2001) was used. The transfer took 1 hour at a current of 200 mA. The transfer buffer consisted of 50 ml of

transfer buffer concentrate (NOVEX NP 0006), 1 ml of antioxidant (NOVEX NP 0002), 100 ml of analytical grade methanol and 849 ml of double-distilled water.

5 Besides the blots produced in this way, also used were premade blots, for example from Chemicon (mouse brain blot, Chemicon, catalog No.: NS 106 with the tissues 1. frontal cortex, 2. posterior cortex, 3. cerebellum, 4. hippocampus, 5. olfactory bulb, 6. striatum, 7. thalamus, 8. mid brain, 9. entorhinal cortex, 10. 10 pons, 11. medulla, 12. spinal cord).

Antibody reaction with PARP3:

The Western blots were blocked in TBST (TBS + 0.3 % Tween 20) 15 with 5% dry milk powder for at least 2 hours (TBS: 100 mM Tris pH 7.5, 200 mm NaCl). The antibody reaction with the primary antibody (dilution 1:1000) took place in TBST with 5% dry milk powder (see above) at room temperature for at least 2 hours or at 4°C overnight, with gentle agitation (vertical rotator). This was 20 followed by washing three times in TBST for 5 minutes. Incubation with the secondary antibody (anti-rabbit IgG, peroxidase-coupled, SIGMA A-6154, dilution 1:2000) took place in TBST with 5% dry milk powder for 1 hour. This was followed by washing three times for 5 minutes each time as above. The subsequent detection was 25 based on chemiluminescence using the SUPER BLAZE kit (Pierce, Signal BLAZE Chemiluminescent Substrate 34095) as stated by the manufacturer. The "Lumi-Film" (Chemiluminescent Detection Film, Boehringer order No: 1666916) was used. The films were developed for about 2 min (X-ray developer concentrate, ADEFO-Chemie GmbH), 30 hydrated, fixed for about 4 min (Acidofix 85 g/l /AGFA), hydrated and then dried.

Example 5: Preparation of the enzymes

35 For comparison, human PARP1 was expressed recombinantly in the baculovirus system in the manner familiar to the skilled worker and partially purified as described (Shah et al., Analytical Biochemistry 1995, 227, 1-13). Bovine PARP1 in a purity of 30-50% (C= 0.22 mg/ml, spec. activity 170 nmol of ADP-ribose/min/mg of total protein at 25°C) was purchased from BIOMOL (order No. SE-165). Human and mouse PARP2 and PARP3 were expressed recombinantly in the baculovirus system (Bac-to-Bac system, BRL LifeScience). For this purpose, the appropriate cDNAs were cloned to the pFASTBAC-1 vector. Preparation of recombinant baculovirus DNA by recombination in E. coli was followed by transfection of insect cells (Sf9 or High-Five) with the appropriate recombinant baculovirus DNAs. Expression of the corresponding proteins was veri-

fied by Western blot analysis. Virus strains were amplified in the manner familiar to the skilled worker. Larger amounts of recombinant proteins were obtained by infecting 500 ml of insect cell culture (2 x 10⁶ cells/ml) with viruses in an MOI (multiplicity of infection; ratio of viruses to cells) of 5-10 and incubating for 3 to 4 days. The insect cells were then pelleted by centrifugation, and the proteins were purified from the pellet.

The purification took place by classical methods of protein puri10 fication familiar to the skilled worker, detecting the enzymes
with appropriate specific antibodies. In some cases, the proteins
were also affinity-purified on a 3-aminobenzamide affinity column
as described (Burtscher et al., Anal Biochem 1986, 152:285-290).
The purity was >90%.

15

Example 6: Assay systems for determining the activity of PARP2 and PARP3 and the inhibitory action of effectors on PARP1, PARP2 and PARP3.

20 a) Production of antibodies against poly(ADP-ribose)

It is possible to use poly(ADP-ribose) as antigen for generating anti-poly(ADP-ribose) antibodies. The production of antipoly(ADP-ribose) antibodies is described in the literature (Kanai Y et al. (1974) Biochem Biophys Res Comm 59:1, 300-306; Kawamaitsu H et al. (1984) Biochemistry 23, 3771-3777; Kanai Y et al. (1978) Immunology 34, 501-508).

The following were used, inter alia: anti-poly(ADP-ribose) anti-30 bodies (polyclonal antiserum, rabbits), BIOMOL; order No. SA-276, anti-poly(ADP-ribose) antibodies (monoclonal, mouse; clone 10H; hybridoma supernatant, affinity-purified).

The antisera or monoclonal antibodies obtained from hybridoma 35 supernatant were purified by protein A affinity chromatography in the manner familiar to the skilled worker.

- b) ELISA
- 40 Materials:

ELISA color reagent: TMB mix, SIGMA T-8540

A 96-well microtiter plate (FALCON Micro-Test III™ Flexible Assay 45 Plate, # 3912) was coated with histones (SIGMA, H-7755). Histones were for this purpose dissolved in carbonate buffer (0.05M Na₂HCO₃; pH 9.4) in a concentration of 50 µg/ml. The individual

wells of the microtiter plate were each incubated with 150 μl of this histone solution at room temperature for at least 2 hours or at 4°C overnight. The wells are then blocked by adding 150 μl of a 1% BSA solution (SIGMA, A-7888) in carbonate buffer at room temperature for 2 hours. This is followed by three washing steps with washing buffer (0.05% Tween10 in 1x PBS; PBS (Phosphate buffered saline; Gibco, order No. 10010): 0.21g/l KH₂PO₄, 9g/l NaCl, 0.726g/l Na₂HPO₄ · 7H₂O, pH 7.4). Washing steps were all carried out in a microtiter plate washer ("Columbus" microtiter plate 10 washer, SLT-Labinstruments, Austria).

Required for the enzyme reaction were an enzyme reaction solution and a substrate solution, in each case as a premix. The absolute amount of these solutions depended on the intended number of 15 assay wells.

Composition of the enzyme reaction solution per well: $-4~\mu l$ of PARP reaction buffer (1M Tris-HCl pH 8.0, 100mM MgCl₂, 10mM DTT)

- 20 20ng of PARP1 (human or bovine) or 8ng PARP2 (human or mouse)
 - 4 μ l of activated DNA (1 mg/ml; SIGMA, D-4522)
 - H_2O ad 40 μ l

Composition of the substrate solution per well:

- 25 5 μ l of PARP reaction buffer (10x)
 - 0.8 μ l of NAD solution (10mM, SIGMA N-1511)
 - $-44 \mu l H_2O$

Inhibitors were dissolved in 1x PARP reaction buffer. DMSO, which 30 was occasionally used to dissolve inhibitors in higher concentrations, was no problem up to a final concentration of 2%. For the enzyme reaction, 40 μ l of the enzyme reaction solution were introduced into each well and incubated with 10 μ l of inhibitor solution for 10 minutes. The enzyme reaction was then 35 started by adding 50 μ l of substrate solution per well. The reaction was carried out at room temperature for 30 minutes and then stopped by washing three times with washing buffer.

The primary antibodies employed were specific anti-poly(ADP-ri40 bose) antibodies in a dilution of 1:5000. Dilution took place in
antibody buffer (1% BSA in PBS; 0.05% Tween20). The incubation
time for the primary antibodies was one hour at room temperature.
After subsequently washing three times with washing buffer, incubation was carried out with the secondary antibody (anti-mouse
45 IgG, Fab fragments, peroxidase-coupled, Boehringer Mannheim,
order No. 1500.686; anti-rabbit IgG, peroxidase-coupled, SIGMA,
order No. A-6154) in a dilution of 1:10,000 in antibody buffer at

room temperature for one hour. Washing three times with washing buffer was followed by the color reaction using 100 μ l of color reagent (TMB mix, SIGMA) per well at room temperature for about 15 min. The color reaction was stopped by adding 100 μ l of 2M 5 H₂SO₄. This was followed by immediate measurement in an ELISA plate reader (EAR340AT "Easy Reader", SLT-Labinstruments, Austria) (450nm versus 620nm). The measurement principle is depicted diagrammatically in Figure 6.

- 10 Various concentrations were used to construct a dose-effect plot to determine the K_i value of an inhibitor. Values are obtained in triplicate for a particular inhibitor concentration. Arithmetic means are determined using Microsoft© Excel. The IC₅₀ is determined using the Microcal© Origin Software (Vers. 5.0)
 15 ("Sigmoidal Fit"). Conversion of the IC₅₀ value is calculated in this way into K_i values took place by using "calibration inhibitors". The "calibration inhibitors" were also measured in each analysis. The K_i values of the "calibration inhibitors" were determined in the same assay system by analysis of the Dixon dia20 gram in the manner familiar to the skilled worker.
 - b) HTRF (homogenous time-resolved fluorescence) assay
- In the HTRF PARP assay according to the invention, histones, as

 25 target proteins for modification by PARP, are labeled indirectly with an XL665 fluorophore. The anti poly(ADP ribose) antibody is directly labeled with a europium cryptate (anti-PAR-cryptate). If the XL665 fluorophore is in the direct vicinity in space, which is ensured by binding to the poly(ADP-ribose) on the histone,

 30 then energy transfer is possible. The emission at 665 nm is thus directly proportional to the amount of bound antibody, which in turn is equivalent to the amount of poly(ADP-ribose). The measured signal thus corresponds to the PARP activity. The measurement principle is depicted diagrammatically in Figure 7.

 35 The materials used are identical to those used in the ELISA (see above) unless expressly indicated.

Histones were dissolved in a concentration of 3 mg/ml in Hepes buffer (50mM, pH=7.5). Biotinylation took place with 40 sulfo-NHS-LC-biotin (Pierce, #21335T). A molar ratio of 4 biotin molecules per histone was used. The incubation time was 90 minutes (RT). The biotinylated histones were then purified on a G25 SF HR10/10 column (Pharmacia, 17-0591-01) in Hepes buffer (50mM, pH=7.0) in order to remove excess biotinylation reagent.

45 The anti-poly(ADP-ribose) antibody was labeled with europium

cryptate using bifunctional coupling reagents (Lopez, E. et al., Clin. Chem. 39(2), 196-201 (1993); US Patent 5,534,622).

Purification took place on a G25SF HR10/30 column. A molar ratio of 3.1 cryptates per antibody was achieved. The yield was 25%. The conjugates were stored at -80° C in the presence of 0.1% BSA in phosphate buffer (0.1M, pH=7).

5

For the enzyme reaction, the following were pipetted into each well:

- 10 μl of PARP solution in PARP HTRF reaction buffer (50mM TrisHCl pH 8.0, 10mM MgCl2, 1mM DTT) with 20ng of PARP1 (human or bov-10 ine) or 8ng of PARP2 (human or mouse)
 - 10 μ l of activated DNA in PARP HTRF reaction buffer (50 μ g/ml)
 - 10 μl of biotinylated histones in PARP HTRF reaction buffer (1.25 μM)
 - 10 μ l of inhibitor in PARP HTRF reaction buffer

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These reagents were incubated for 2 minutes before the reaction was started by adding

- 10 μl of NAD solution in PARP HTRF reaction buffer (41 $\mu M/m l)$. The reaction time was 30 minutes at room temperature.

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The reaction was then stopped by adding

- 10 μ l of PARP inhibitor (25 μ M, K_i =10nM) in "Revelation" buffer (100mM Tris-HCl pH 7.2, 0.2M KF, 0.05% BSA).
- 25 The following were then added:
 - 10 μ l of EDTA solution (SIGMA, E-7889, 0.5M in H₂O)
 - 100 μ l of Sa-XL665 (Packard Instruments) in "Revelation" buffer (15-31.25nM)
 - 50 μ l of anti-PAR cryptate in "Revelation" buffer (1.6-3.3nM).

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Measurement was then possible after 30 minutes (up to 4 hours). The measurement took place in a "discovery HTRF microplate analyzer" (Canberra Packard Instruments). The K_i values were calculated as described for the ELISA.

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Example 7: Test systems for determining the therapeutic efficacy of PARP inhibitors

40 Novel PARP inhibitors can have their therapeutic efficacy checked in relevant pharmacological models. Examples of some suitable models are listed in Table 1.

	Disorder	Model	Literature				
5	Neurodegenerative disorders (stroke, Parkinson's, etc.)	NMDA excitotoxicity in mice or rats	See below for des- cription				
10	Stroke	Permanent MCAO ("middle cerebral arterial occlusion")	Tokime, T. et al., J. Cereb. Blood Flow Metab., 18(9): 991-7, 1998. Guegan, C., Brain Research. Molecular Brain Research, 55(1): 133-40, 1998.				
15		Transient, focal MCAO in rats or mice	Eliasson MJL et al., Nat Med 1997, 3:1089-1095.				
13			Endres, M et al., J Cereb Blood Flow Metab 1997, 17:1143-1151.				
20			Takahashi K et al., J Cereb Blood Flow Metab 1997, 17:1137-1142.				
25	Parkinson's disease	MPTP (1-methyl- 4-phenyl-1,2,3,6- tetrahydropyridine)	Cosi C, et al., Brain Res., 1998 809(1):58-67.				
		toxicity in mice/ rats	Cosi C, et al., Brain Res., 1996 729(2):264-9.				
30	Myocardial infarct	Coronary vessel occlusion in rats, pigs or rabbits	Richard V, et al., Br. J. Pharmacol 1994, 113, 869-876.				
			Thiemermann C, et al., Proc Natl Acad Sci U S A. 1997, 94(2):679-83.				
35			Zingarelli B, et al., Cardiovasc Res. 1997, 36(2):205-15.				
40		Langendorf heart model in rats or rabbits	See below for des- cription				
	Septic shock	Endotoxin shock in rats	Szabo C, et al., J Clin Invest, 1997, 100(3):723-35.				

5		Zymosan- or carrageenan-induced multiple organ failure in rats or mice	Szabo C, et al. J Exp Med. 1997, 186(7):1041-9.Cuzzo- crea S, et al. Eur J Pharmacol. 1998, 342(1):67-76.
	Rheumatoid arthritis	Adjuvant- or collagen-induced arthritis in rats or mice	Szabo C, et al., Proc Natl Acad Sci U S A. 1998, 95(7):3867-72.
10	Diabetes	Streptozotocin- and alloxan-induced or obesity-associated	Uchgata Y et al., Diabetes 1983, 32: 316-318.Masiello P et al., Diabetologia 1985, 28: 683-686.Shimabukuro M et al., J Clin In- vest 1997, 100: 290-295.
	Cancer	In vitro model; see below	Schlicker et al., 1999, 75(1), 91-100.

20

a) NMDA excitotoxicity model

Glutamate is the most important excitory neurotransmitter in the brain. Under normal conditions, glutamate is secreted into the synaptic cleft and stimulates the post-synaptic glutamate receptors, specifically the glutamate receptors of the "NMDA" and "AMPA" types. This stimulation plays a significant part in numerous functions of the brain, including learning, memory and motor control.

30

Under the conditions of acute and chronic neurodegeneration (e.g. stroke), however, there is a great increase in the presynaptic glutamate secretion, resulting in excessive stimulation of the receptors. This leads to death of the cells stimulated in this 35 way. These increased glutamate activities occur in a number of neurological disorders or psychological disturbances and lead to states of overexcitation or toxic effects in the central nervous system (CNS) but also in the peripheral nervous system. Thus, glutamate is involved in a large number of neurodegenerative dis-40 orders, in particular neurotoxic disturbances following hypoxia, anoxia, ischemia and after lesions like those occurring after stroke and trauma, and stroke, Alzheimer's disease, Huntington's disease, amyotrophic lateral sclerosis (ALS; "Lou Gehring's disease"), cranial trauma, spinal cord trauma, peripheral neuro-45 pathies, AIDS dementia and Parkinson's disease. Another disease in which glutamate receptors are important is epilepsy (cf. Brain Res Bull 1998; 46(4):281-309, Eur Neuropsychopharmacol 1998, 8(2):141-52.).

Glutamate effects are mediated through various receptors. One of these receptors is called the NMDA (N-methyl-D-aspartate) receptor after a specific agonist (Arzneim.Forschung 1990, 40, 511-514; TIPS, 1990, 11, 334-338; Drugs of the Future 1989, 14, 1059-1071). N-Methyl-D-aspartate is a strong agonist of a particular class of glutamate receptors ("NMDA" type). Stimulation of the NMDA receptor leads to influx of calcium into the cell and the generation of free radicals. The free radicals lead to DNA damage and activation of PARP. PARP in turn causes cell death through depletion of high-energy phosphates (NAD and ATP) in the cell. This explains the toxicity of NMDA. Treatment of animals with NMDA can therefore be regarded as a model of the abovementioned disorders in which excitotoxicity is involved.

Because of the importance of glutamate receptors in neurodegeneration, many pharmacological approaches to date have been directed 20 at specific blocking of precisely these receptors. However, because of their importance in normal stimulus conduction, these approaches have proved to be problematic (side effects). In addition, stimulation of the receptors is an event which takes place very rapidly so that administration of the receptors often comes too late ("time window" problem). Thus there is a great need for novel principles of action and inhibitors of NMDA-related neurotoxicity.

Protection against cerebral overexcitation by excitatory amino 30 acids (NMDA antagonism in mice) can be regarded as adequate proof of the activity of a pharmacological effector of PARP in disorders based on excitotoxicity. Intracerebral administration of excitatory amino acids (EAA) induces such massive overexcitation that it leads within a short time to convulsions and death of the 35 animals (mice).

In the present case there was unilateral intracerebroventricular administration of 10 µl of a 0.035% strength aqueous NMDA solution 120 minutes after intraperitoneal (i.p.) administration of the 40 test substance. These symptoms can be inhibited by systemic, e.g. intraperitoneal, administration of centrally acting drugs. Since excessive activation of EAA receptors in the central nervous system plays an important part in the pathogenesis of various neurological disorders, information can be gained from the 45 detected EAA antagonism in vivo about possible therapeutic utilizability of the substances for such CNS disorders. An ED50 at which 50% of the animals are, due to preceding i.p.

administration of the measured substance, free of symptoms with a fixed dose of NMDA was determined as a measure of the activity of the substances.

5 b) Langendorf heart model (model for myocardial infarct)

Male Sprague-Dawley rats (bodyweight 300-400 g; origin Janvier, Le Genest-St-Isle, France) were used for the test. The rats were treated orally by gavage with the active substance or placebo 10 (volume: 5 ml/kg). 50 minutes later, heparin is administered intraperitoneally (Liquemin N Roche, 125 IU/animal in 0.5 ml). The animals are anesthesized with Inactin® T133 (thiobetabarbital sodium 10%), fixed on the operating table, tracheotomized and ventilated with a "Harvard ventilatory pump" (40 beats/min, 15 4.5 ml/beat). Thoracotomy was followed by immediate catheterization of the aorta, removal of the heart and immediate retrograde perfusion. The hearts were perfused with a constant pressure of 75 mmHg, which is achieved using a "Gilson Miniplus 2 perfusion pump". Composition of the perfusate (mmol/1): NaCl 118, KCl 4.7, **20** CaCl₂ x 2 H₂O 2.52, MgSO₄ x 7 H₂O 1.64, NaHCO₃ 24.88, KH₂PO₄ 1.18, glucose 11. The temperature is kept at 37°C throughout the experiment. Functional parameters were continuously recorded using a "Gould 4-channel recorder". Measurements were made of the leftventricular pressure (LVP; mmHg), LVEDP (mmHg), enzyme release 25 (creatine kinase, mU/ml/g), coronary flow rate (ml/min), HR (pulse rate, min-1). The left-ventricular pressure was measured using a liquid-filled latex balloon and a Statham23 Db pressure transducer. The volume of the balloon was initially adjusted to reach an LVEDP (left-ventricular end-diastolic pressure) of about 30 12 mmHg. The dP/dt_{max} (maximum pumping force) is derived from the pressure signal using a differentiator module. The heart rate was calculated from the pressure signal. The flow rate was determined using a drop counter (BMT Messtechnik GmbH Berlin). After an equilibration time of 20 minutes, the hearts were subjected to a 35 30-minute global ischemia by stopping the perfusate supply while keeping the temperature at 37°C. During the following 60-minute reperfusion period, samples of the perfusate were taken after 3, 5, 10, 15, 30, 45 and 60 min for analysis of creatine kinase (CK) activity. Means and standard deviations for the measured para-40 meters were analyzed statistically (Dunnett test). The signifi-

The experiment on rabbit hearts was carried out similarly. Male white New Zealand rabbits (obtained from: Interfauna) were used.

45 The hearts were prepared as described above for the rat model. The perfusion pressure was set at a maximum of 60 mmHg and the flow rate at about 25ml/min. The equilibration time was about

cance limit was p=0.05.

30 min. The substance was administered by infusion directly upstream of the heart. 15 min after starting the infusion, a 30-minute global ischemia was caused by stopping the flow while maintaining the temperature of the heart. A 30-minute reperfusion 5 followed. Perfusate was taken for investigation of CK activity before administration of the substance, after 15 min and at various times (5, 10, 15, 20, 30 min) during the reperfusion. The following parameters were measured: LVP (mmHg), LVEDP, LVdP/dt, PP (mmHg), HR (pulse rate; beats/min), CK activity (U/min/g heart 10 weight).

c) Animal model for acute kidney failure

The protective effect of intravenous administration of PARP 15 inhibitors (4 days) on the kidney function of rats with postischemic acute kidney failure was investigated.

Male Sprague-Dawley rats (about 330 g at the start of the experiments; breeder: Charles River) were used. 10-15 animals were 20 employed per experimental group. Administration of active substance/placebo took place continuously with an osmotic micropump into the femoral vein. Orbital blood was taken (1.5 ml of whole blood) under inhalation anesthesia with enflurane (Ethrane Abbot, Wiesbaden).

25 After the initial measurements (blood sample) and determination of the amount of urine excreted in 24h, the rats were anesthetized ("Nembutal", pentobarbital sodium, Sanofi CEVA; 50mg/kg i.p., volume injected 1.0 ml/kg) and fastened on a heatable oper-30 ating table (37°C). 125 IU/kg heparin (Liquemin N, Roche) were administered i.v. into the caudal vein. The abdominal cavity was opened and the right kidney was exposed. The branching-off renal artery was exposed and clamped off superiorly using bulldog clamps (Diefenbach 38mm). The left renal artery was likewise ex-35 posed and clamped off (superiorly, about half way to the kidney). During the operation, an osmotic micropump was implanted into the femoral vein. The intestine was reinserted and the fluid loss was compensated with luke-warm 0.9% NaCl. The animals were covered with a moist cloth and kept warm under red light. After 40 min, 40 the appearance of the kidneys was recorded, and the clamps were removed, first the right then the left. The intestine was put back and 2 drops of antibiotic (Tardomyocel, Bayer) were added. The abdominal wall was closed with sterile cat gut (Ethicon No.4) and treated once more with 1 drop of antibiotic. The epidermis 45 was sutured with sterile Ethibond Exel (Ethicon) No.3/0, and the

suture was sprayed with Nebacetin N (Yamanouchi) wound spray. A tenth of a daily dose of drug/placebo is given as i.v. bolus.

Samples and blood were taken for investigating biochemical para5 meters in the serum and urine: Na, K, creatinine, protein (only in urine), on days 1, 2 and 4 of the experiment. In addition, the feed and water consumption, bodyweight and urine volume were recorded. After 14 days, the animals were sacrificed and the kidneys were assessed.

10

The assessment excluded all animals which died of an infarct during the experiment or showed an infarct at necropsy on day 14.

The creatinine clearance and the fractional sodium excretion were calculated as kidney function parameters, comparing treated ani
15 mals with control and sham.

- d) In vitro model for radiosensitization (tumor therapy)
- MCF-7-cells (human breast carcinoma) were cultivated in

 20 Dulbecco's modified Eagle's medium with 10% heat-inactivated FCS and 2 mM L-glutamine. Cells were seeded out overnight in cell densities of 100, 1000 or 10,000 cells per well in a 6-well plate and then exposed to ionizing radiation with a dose in the range from 0 to 10 Gy (137Cs, Shepard Mark, model I-68A, dose rate

 25 3.28 Gy/min). 10 days after the irradiation, the experiment was assessed, counting colonies with fifty cells as positive.
 - e) Stroke model (focal cerebral ischemia; MCA (middle cerebral artery) occlusion on a rat)

30

A focal ischemia was performed by means of cauterisation of the right distal MCA on Sprague-Dawley or Long-Evans rats. The rats may be treated before or after the beginning of the MCA occlusion with modulators of the proteins of the invention. As a rule, 35 doses of 1-10 mg/kg are chosen (bolus application), optionally followed by a continuous infusion of 0.5-5 mg/kg/h.

The rats are anesthetised with halothane in a mixture of 70 % nitrogen and 30 % oxygen (4% at initial phase and 0.8-1.2 % during

- 40 the operation). The body temperature was permanently measured rectally and was kept constant at 37.5 °C \pm 0.5 °C by means of a controllable heating blanket. Moreover, arterial blood pressure, arterial pH, (Pa(O₂) and Pa(CO₂) were optionally measured by means of a tail vein catheder. Thereafter, the focal iscehmia was car-
- 45 ried out using the method of Chen et al. (Stroke 17: 738-743; 1986) or Liu et al. (Am. J. Physiol. 256: H589-593; 1989) by means of continuous cauterisation of the distal part of the right

MCA. When the operation was terminated, the animals were kept in a warm environment for a further 24 hours. Then they were killed with the use of CO₂ and decapitated. Their brains were taken, shock-frozen (dry ice or liquid nitrogen) and stored at -80 °C.

5 The brains were cut into 0.02 mm thick slices and every 20th cut was used for the subsequent analysis. The corresponding cuts are stained with cresyl violet (Nissl staining). Alternatively, TTC (2,3,4-triphenyltetrazoliumchloride) may be used for staining. The infarct volume may then be analysed under a microscope. For 10 exact quantification, a computer-based image analyzing software may be used (J. Cereb. Clood Flow Metabol. 10: 290-293; 1990).

f) Septic shock

- 15 Groups of 10 male C57/BL mice (body weight 18-20 g) are treated with LPS (lipopolysaccharide, from E. coli, LD₁₀₀ 20 mg/animal i. v.) plus galactosamine (20 mg/animal i. v.). the substance to be tested is applied i. p. or i. v. during three succeeding days (e. g. 1-10 mg/kg), with the first dose being administered 30 minutes after the LPS treatment. The death rate is determined every 12 hours. Alternatively, the substance may also be applied in several doses spread over the days.
 - g) Determination of altered gene expression in aging cells
 - The aging of cells is simulated by changing the cell culture media from the complete medium with a reduced serum concentration and thereafter is analysed by means of quantitative PCR or Northern Blotting (Linskens et al., Nucleic Acids Res. 1995, 23(16):
- 30 3244-51). As typical markers for the aging of the skin for example collagen or elastin may be used. Human fibroblasts or fibroblast cell lines are used which simulate the aging of the skin. Modulators of the proteins of the invention are added to the medium and their effect on the changing of the gene express-
- 35 ion is observed. An increased production of elastin in cells with a reduced aging process caused by means of said modulators may be observed.

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SEQUENCE LISTING

- (i) APPLICANT:
 - (A) NAME: BASF Aktiengesellschaft
 - (B) STREET:
 - (C) CITY: Ludwigshafen
 - (E) COUNTRY: Deutschland
 - (F) POSTAL CODE (ZIP): 67065
- (ii) TITLE OF INVENTION: Neue Poly ADP Ribose Polymerase Gene
- (iii) NUMBER OF SEQUENCES: 28
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC DOS/MS DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1843 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (F) TISSUE TYPE: Brain
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 3...1715
 - (D) OTHER INFORMATION:/product= "Poly ADP Ribose Polymerase"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:
- GCA TTA AAT GAA AGC AAA AGA GTT AAT AAT GGC AAC ACG GCT CCA GAA 95
 Ala Leu Asn Glu Ser Lys Arg Val Asn Asn Gly Asn Thr Ala Pro Glu
 20 25 30

TCT Ser								143
AAG Lys								191
AAG Lys 65								239
GTG Val								287
GAA Glu								335
TTC Phe								383
 CAG Gln								431
GGA Gly 145								479
GAA Glu								527
GAT Asp								575
ATG Met								623
GAA Glu								671
GTA Val 225								719
ATG Met								767
CTG Leu								815

	260	265	2	70
			GA CGA GCT CTC A Bly Arg Ala Leu M 285	
	Phe Tyr Thr A		AT GAC TTT GGA C is Asp Phe Gly I 300	
			TG TCA GAA AAA A Leu Ser Glu Lys I 315	
		Ile Glu Ile A	GCT ATT AAG CTG G lla Ile Lys Leu V 330	
			GAC CAA CAC TAT A Asp Gln His Tyr A 3	
			GAA AGT TAC GAG T Glu Ser Tyr Glu F 365	
	Tyr Leu Gln S		GCT CCC ACA CAC A Ala Pro Thr His S 380	
			ETG GAG AAG GAT G 7al Glu Lys Asp G 395	
		Leu His Asn A	AGG ATG CTT CTA T Arg Met Leu Leu T 110	
			TTG AGC CAT GGG C Leu Ser His Gly I	
			TAC ATG TTT GGG A Tyr Met Phe Gly I 445	
	Asp Met Ser S		GCC AAT TAC TGC 1 Ala Asn Tyr Cys I 460	
			TTA TCA GAG GTA (Leu Ser Glu Val <i>I</i> 475	
		Glu Ala Asn F	CCT AAG GCC GAA (Pro Lys Ala Glu (190	
CTT CAA GGT AAA	CAT AGC ACC	AAG GGG CTG G	GGC AAG ATG GCT (CCC AGT 1535

0050/49100/49790

Leu	Gln	Gly	Lys	His 500	Ser	Thr	Lys	Gly	Leu 505	Gly	Lys	Met	Ala	Pro 510	Ser	
				-										GGA Gly	CCA Pro	1583
														AAC Asn		1631
				_										TAC Tyr		1679
	AAG Lys											ATG:	PTGA:	TAT		1725
TAAZ	ATAAI	ACC I	AGAGA	ATCTO	A TO	CTTC	AAGC	A AGA	AAAA	raag	CAG	rgtt	GTA (CTTG	rgaatt	1785
TTG	rgat <i>i</i>	ATT :	TATO	TAAT	CA A	AAAC	rgtac	CAG	STCT	AAAA	AAA	AAAA	AAA .	AAAA	AAAA	1843

(2) INFORMATION FOR SEQ ID NO: 2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 571 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Ala Ala Arg Arg Arg Ser Thr Gly Gly Arg Ala Arg Ala 1 5 10 15

Leu Asn Glu Ser Lys Arg Val Asn Asn Gly Asn Thr Ala Pro Glu Asp 20 25 30

Ser Ser Pro Ala Lys Lys Thr Arg Arg Cys Gln Arg Gln Glu Ser Lys 35 40 45

Lys Met Pro Val Ala Gly Gly Lys Ala Asn Lys Asp Arg Thr Glu Asp 50 55 60

Lys Gln Asp Glu Ser Val Lys Ala Leu Leu Leu Lys Gly Lys Ala Pro 65 70 75 80

Val Asp Pro Glu Cys Thr Ala Lys Val Gly Lys Ala His Val Tyr Cys
85 90 95

Glu Gly Asn Asp Val Tyr Asp Val Met Leu Asn Gln Thr Asn Leu Gln 100 105 110

Phe Asn Asn Asn Lys Tyr Tyr Leu Ile Gln Leu Leu Glu Asp Asp Ala 115 120 125

Gln Arg Asn Phe Ser Val Trp Met Arg Trp Gly Arg Val Gly Lys Met

	130					135					140				
Gly 145	Gln	His	Ser	Leu	Val 150	Ala	Cys	Ser	Gly	Asn 155	Leu	Asn	Lys	Ala	Lys 160
Glu	Ile	Phe	Gln	Lys 165	Lys	Phe	Leu	Asp	Lys 170	Thr	Lys	Asn	Asn	Trp 175	Glu
Asp	Arg	Glu	Lys 180	Phe	Glu	Lys	Val	Pro 185	Gly	Lys	Tyr	Asp	Met 190	Leu	Gln
Met	Asp	Tyr 195	Ala	Thr	Asn	Thr	Gln 200	Asp	Glu	Glu	Glu	Thr 205	Lys	ГÀЗ	Glu
Glu	Ser 210	Leu	Lys	Ser	Pro	Leu 215	Lys	Pro	Glu	Ser	Gln 220	Leu	Asp	Leu	Arg
225					230					235			Met		240
				245					250				Leu	255	
			260					265					Lys 270		
,	_	275					280					285	Met		
-	290			_		295					300		Leu		
305					310					315			Ile		320
		-		325					330				Val	335	
			340					345					Arg 350		
	_	355					360					365	Phe		
	370					375					380		Ser		
385					390					395			Gly		400
				405					410				Trp	415	
			420					425					Leu 430		
		435					440					445	Lys		
Tyr	Phe	Ala	Asp	Met	Ser	Ser	Lys	Ser	Ala	Asn	Tyr	Cys	Phe	Ala	Ser

	450					455					460					
Arg 465	Leu	Lys	Asn	Thr	Gly 470	Leu	Leu	Leu	Leu	Ser 475	Glu	Val	Ala	Leu	Gly 480	
Gln	Cys	Asn	Glu	Leu 485	Leu	Glu	Ala	Asn	Pro 490	Lys	Ala	Glu	Gly	Leu 495	Leu	
Gln	Gly	Lys	His 500	Ser	Thr	Lys	Gly	Leu 505	Gly	Lys	Met	Ala	Pro 510	Ser	Ser	
Ala	His	Phe 515	Val	Thr	Leu	Asn	Gly 520	Ser	Thr	Val	Pro	Leu 525	Gly	Pro	Ala	
Ser	Asp 530	Thr	Gly	Ile	Leu	Asn 535	Pro	Asp	Gly	Tyr	Thr 540	Leu	Asn	Tyr	Asn	
Glu 545	Tyr	Ile	Val	Tyr	Asn 550	Pro	Asn	Gln	Val	Arg 555	Met	Arg	Tyr	Leu	Leu 560	
Lys	Val	Gln	Phe	Asn 565	Phe	Leu	Gln	Leu	Trp 570	*						
(2)	INF	ORMA	rion	FOR	SEQ	ID i	NO: 3	3:								
	<pre>(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2265 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI SENSE: NO (vi) ORIGINAL SOURCE: (F) TISSUE TYPE: Uterus (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION:2421843 (D) OTHER INFORMATION:/product= "Poly ADP Ribose Polymerase"</pre>															
тсс	•	•	-				ON:					rece:	ልሮሞ (CCTC	GCCAAA	<u> </u>
															GAATTO	
TCT	CCCT	AAT '	TCAC(GCCT	GA G	GCTC	ATGG	A GA	GTTG	CTAG	ACC'	rggg	ACT (GCCC'	rggga	180
GCG	CACA	CAA (CCAG	GCCG	GG T	GGCA	GCCA	G GA	CCTC'	TCCC	ATG'	rccc'	rgc '	TTTT	CTTGGC	240
	TG G													CT G		286

	575							58	30							
												CCC Pro				334
												CGC Arg 615				382
												ACC Thr				430
												GAG Glu				478
												AAC Asn				526
												GGC Gly				574
												GAC Asp 695				622
												CGG Arg				670
												CAG Gln				718
												CCA Pro				766
												GCC Ala				814
			Asn									AAC Asn 775			GCC Ala	. 862
												AAG Lys				910
CAA Gln 795	Gln	ATT Ile	GCA Ala	CGG Arg	GGT Gly 800	TTC Phe	GAG Glu	GCC Ala	TTG Leu	GAG Glu 805	GCG Ala	CTG Leu	GAG Glu	GAG Glu	GCC Ala 810	958
CTG	AAA	GGC	CCC	ACG	GAT	GGT	GGC	CAA	AGC	CTG	GAG	GAG	CTG	TCC	TCA	1006

Leu Lys Gly	Pro Thr 815	Asp Gly G	ly Gln Ser 820		Glu Leu Ser 825	Ser
CAC TTT TAC						
CCC ATC AAS Pro Ile Ass 845	Ser Pro	Glu Leu L		Lys Lys A		
GTG CTG GCC Val Leu Ala 860						
CAG GAG AAG Gln Glu Ly: 875						
CAG CTT CTO				Asp Ser (GGA GCA CCT Gly Ala Pro 905	
					AGC AAC CAC Ser Asn His 920	
	Leu Gln	His Ile T		Asn Gln (GAA GGG GAG Glu Gly Glu 935	
					AAG CTG CTG Lys Leu Leu	
					ACT AGT GGG Thr Ser Gly	
				. Gly Lys (GGC ATC TAC Gly Ile Tyr 985	Phe
					GGC ATG AAG Gly Met Lys 1000	
	s His Val	Gly Tyr M		ı Gly Glu '	GTG GCC CTG Val Ala Leu 1015	
					AAG AGC CCA Lys Ser Pro	
					GAG CCT GAT Glu Pro Asp	

														GTG Val		1726
1111	GIII	изр		1055		GIU	neu	1155	1060		0 2 · · ·	• • • •	• • • •	1065		
														TTC		1774
GIn	GLY	Gin	1070		Pro	Cys	PIO	1075		ser	ser	ser	1080	Phe)	ser	
						_	-							CGC		1822
Gln	Ser	Glu 1085	-	Leu	Ile	Tyr	1090		Ser	Gln	Cys	Arg 1099		Arg	Tyr	
							GTGC	CCGC	CC 1	GTC	cccc	GG GG	STCC	rgcaa	Δ.	1873
Leu	Leu 1100		Val	His	Leu	1105	5									
GGC1	rggac	TG T	rgato	CTTC	AA TO	CATCO	CTGCC	CAT	CTCI	GGT	ACC	CTA	TAT (CACTO	CCTTTT	1933
TTTC	CAAGA	AT A	ACAA	racgi	rt Gi	TGT	TAACI	ATA	AGTC	ACCA	TGCT	CTAC	CAA (GATCO	CCTGAA	1993
CTT	ATGC	CTC (CTAAC	CTGA	AA T	TTGT	TATTO	TTT	GAC	ACAT	CTG	CCA	GTC (CCTCI	CCTCC	2053
CAG	CCAT	rgg :	raaco	CAGC	AT T	rgac:	CTT	AC	TGT	AATA	GGG	CAGC	rtt 1	TATAC	GTTCC	2113
ACA:	rgta <i>i</i>	AGT (GAGA:	CATO	GC A	GTGTT	TGT	TTT	CTGT	rgcc	TGG	CTTA	CTT (CACTO	CAGCAT	2173
AAT	STGC	ACC (GGT:	CAC	CC AS	rgtti	TCAT	' AA	ATGAC	CAAG	ATT	CCT	CCT S	TTAA?	AAAAA	2233
AAA	AAAA	AAA A	AAAA	AAAA	AA AA	AAAA	AAAA	AA A								2265

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 534 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Ala Pro Lys Pro Lys Pro Trp Val Gln Thr Glu Gly Pro Glu Lys
1 10 15

Lys Lys Gly Arg Gln Ala Gly Arg Glu Glu Asp Pro Phe Arg Ser Thr 20 25 30

Ala Glu Ala Leu Lys Ala Ile Pro Ala Glu Lys Arg Ile Ile Arg Val

Asp Pro Thr Cys Pro Leu Ser Ser Asn Pro Gly Thr Gln Val Tyr Glu
50 55 60

Asp Tyr Asn Cys Thr Leu Asn Gln Thr Asn Ile Glu Asn Asn Asn 65 70 75 80

Lys Phe Tyr Ile Ile Gln Leu Leu Gln Asp Ser Asn Arg Phe Phe Thr 85 90 95

Cys Trp Asn Arg Trp Gly Arg Val Gly Glu Val Gly Gln Ser Lys Ile

			100					105					110		
Asn	His	Phe 115	Thr	Arg	Leu	Glu	Asp 120	Ala	Lys	Lys	Asp	Phe 125	Glu	Lys	Lys
Phe	Arg 130	Glu	Lys	Thr	Lys	Asn 135	Asn	Trp	Ala	Glu	Arg 140	Asp	His	Phe	Val
Ser 145	His	Pro	Gly	Lys	Tyr 150	Thr	Leu	Ile	Glu	Val 155	Gln	Ala	Glu	Asp	Glu 160
Ala	Gln	Glu	Ala	Val 165	Val	Lys	Val	Asp	Arg 170	Gly	Pro	Val	Arg	Thr 175	Val
Thr	Lys	Arg	Val 180	Gln	Pro	Cys	Ser	Leu 185	Asp	Pro	Ala	Thr	Gln 190	Lys	Leu
Ile	Thr	Asn 195	Ile	Phe	Ser	Lys	Glu 200	Met	Phe	Lys	Asn	Thr 205	Met	Ala	Leu
Met	Asp 210	Leu	Asp	Val	Lys	Lys 215	Met	Pro	Leu	Gly	Lys 220	Leu	Ser	Lys	Gln
Gln 225	Ile	Ala	Arg	Gly	Phe 230	Glu	Ala	Leu	Glu	Ala 235	Leu	Glu	Glu	Ala	Leu 240
Lys	Gly	Pro	Thr	Asp 245	Gly	Gly	Gln	Ser	Leu 250	Glu	Glu	Leu	Ser	Ser 255	His
Phe	Tyr	Thr	Val 260	Ile	Pro	His	Asn	Phe 265	Gly	His	Ser	Gln	Pro 270	Pro	Pro
Ile	Asn	Ser 275	Pro	Glu	Leu	Leu	Gln 280	Ala	Lys	Lys	Asp	Met 285	Leu	Leu	Val
Leu	Ala 290	Asp	Ile	Glu	Leu	Ala 295	Gln	Ala	Leu	Gln	Ala 300	Val	Ser	Glu	Gln
Glu 305	Lys	Thr	Val	Glu	Glu 310	Val	Pro	His	Pro	Leu 315	Asp	Arg	Asp	Tyr	Gln 320
Leu	Leu	Lys	Cys	Gln 325	Leu	Gln	Leu	Leu	Asp 330	Ser	Gly	Ala	Pro	Glu 335	Tyr
Lys	Val	Ile	Gln 340	Thr	Tyr	Leu	Glu	Gln 345	Thr	Gly	Ser	Asn	His 350	Arg	Cys
Pro	Thr	Leu 355	Gln	His	Ile	Trp	Lys 360	Val	Asn	Gln	Glu	Gly 365	Glu	Glu	Asp
Arg	Phe 370	Gln	Ala	His	Ser	Lys 375		Gly	Asn	Arg	Lys 380	Leu	Leu	Trp	His
Gly 385		Asn	Met	Ala	Val 390	Val	Ala	Ala	Ile	Leu 395		Ser	Gly	Leu	Arg 400
Ile	Met	Pro	His	Ser 405		Gly	Arg	Val	Gly 410		Gly	Ile	Tyr	Phe 415	
Ser	Glu	Asn	Ser	Lvs	Ser	Ala	Glv	Tyr	Val	Ile	Gly	Met	Lys	Cys	Gly

			420					425					430			
Ala	His	His 435	Val	Gly	Tyr	Met	Phe 440	Leu	Gly	Glu	Val	Ala 445	Leu	Gly	Arg	
Glu	His 450	His	Ile	Asn	Thr	Asp 455	Asn	Pro	Ser	Leu	Lys 460	Ser	Pro	Pro	Pro	
Gly 465	Phe	Asp	Ser	Val	Ile 470	Ala	Arg	Gly	His	Thr 475	Glu	Pro	Asp	Pro	Thr 480	
Gln	Asp	Thr	Glu	Leu 485	Glu	Leu	Asp	Gly	Gln 490	Gln	Val	Val	Val	Pro 495	Gln	
Gly	Gln	Pro	Val 500	Pro	Cys	Pro	Glu	Phe 505	Ser	Ser	Ser	Thr	Phe 510	Ser	Gln	
Ser	Glu	Tyr 515	Leu	Ile	Tyr	Gln	Glu 520	Ser	Gln	Cys	Arg	Leu 525	Arg	Tyr	Leu	
Leu	Glu 530	Val	His	Leu	*											
	(i)	(I (I	A) LI B) T: C) S:	CE CI ENGTI YPE: FRANI OPOLO	nuci nuci DEDNI	265 l Leic ESS:	ase acio sino	pai: d	cs							
	•			LE T			A									
	(iii)			ETICA ENSE		10										
	•	OR:	IGIN	AL SO	OURCI		Jteri	us								
	(ix)	(1	A) NA B) L(AME/I OCAT: THER	ION:	221. ORMA	rion		oduc	t= "]	Poly	ADP	Rib	ose		
	(xi) SE	QUEN	CE D	ESCR:	IPTI	ON:	SEQ :	ID N	D: 5	:					
TGG	GACT	GGT (CGCC'	rgac'	rc G	GCCT	GCCC	C AG	CCTC'	rgct	TCA	ccc	ACT (GGTG	GCCAAA	60
TAG	CCGA!	rgt (CTAA'	rccc	CC A	CACA	AGCT	C AT	CCCC	GGCC	TCT	GGGA'	rtg '	TTGG(GAATTC	120
TCT	CCCT	AAT '	CAC	GCCT	GA G	GCTC	ATGG	A GA	GTTG	CTAG	ACC'	rggg	ACT (GCCC'	TGGGAG	180
GCG	CACA	CAA (CCAG	GCCG	GG T	GGCA(GCCA	G GA	CCTC'	rccc					TTC Phe	235
TTG	GCC	ATG	GCT	CCA	AAG	CCG	AAG	CCC	TGG	GTA	CAG	ACT	GAG	GGC	CCT	283

Leu 540	Ala	Met	Ala	Pro	Lys 545	Pro	Lys	Pro	Trp	Val 550	Gln	Thr	Glu	Gly	Pro 555	
-		AAG Lys														331
		GCT Ala														379
		GAT Asp 590														427
		GAC Asp														475
		AAG Lys														523
		TGC Cys														571
		AAC Asn														619
		TTT Phe 670														667
		TCT Ser														715
		GCC Ala														763
		ACT Thr														811
		ATC Ile														859
		ATG Met 750														907
		CAG Gln														955

	GGC Gly							1003
	TAC Tyr							1051
	AAT Asn 815							1099
	GCG Ala							1147
 	AAG Lys							1195
	CTC Leu							1243
	GTG Val							1291
	ACA Thr 895							1339
	TTC Phe							1387
	ACC Thr							1435
	ATG Met							1483
	GAG Glu							1531
 	 CAC His 975							1579
	CAC His					Lys		1627
	TTC Phe							1675

	1005	5				1010)				1015	j .				
	Thr					Leu					Gln			GTG Val	GTG Val 1035	1723
					Val					Phe				ACA Thr 1050	Phe	1771
				Tyr					${\tt Glu}$						CGC Arg	1819
			Glu						CCGG	ecc 1	rgtc(cccc	GG G	GTCCI	rgcaa	1873
GGC:	rgga	CTG :	rgat(CTTC	AA T	CATC	CTGC	CA	rctc:	rggt	ACC	CCTAT	PAT (CACTO	CCTTTT	1933
TTT	CAAG	AAT I	ACAA'	racg:	rT G	rtgt:	raac:	r ata	AGTC	ACCA	TGC	rgtac	CAA (GATC	CCTGAA	1993
CTT	ATGC	CTC (CTAA	CTGA	AA T'	TTTG:	TATTO	C TT	rgaci	ACAT	CTG	CCA	GTC (CCTC	rcctcc	2053
CAG	CCCA	rgg '	TAAC	CAGC	AT T'	rgac'	rctt:	r ac'	rtgt/	ATAA	GGG	CAGC!	rtt '	TATA	GGTTCC	2113
ACA'	rgta.	AGT (GAGA'	TCAT	GC A	GTGT'	rtgt(C TT	rctg'	rgcc	TGG	CTTA:	TTT ·	CACT	CAGCAT	2173
AAT	GTGC	ACC (GGGT'	TCAC	CC A	rg t t'	rtca:	r AA	ATGA	CAAG	ATT	rccr	CCT	TTAA	AAAAA	2233
AAA	AAAA	AAA .	AAAA	AAAA	AA A.	AAAA	AAAA	A AA								2265
(2)	INF	ORMA	TION	FOR	SEQ	ID 1	NO: (6:								

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 541 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Ser Leu Leu Phe Leu Ala Met Ala Pro Lys Pro Lys Pro Trp Val 1 5 10 15

Gln Thr Glu Gly Pro Glu Lys Lys Gly Arg Gln Ala Gly Arg Glu 20 25 30

Glu Asp Pro Phe Arg Ser Thr Ala Glu Ala Leu Lys Ala Ile Pro Ala 35 40 45

Glu Lys Arg Ile Ile Arg Val Asp Pro Thr Cys Pro Leu Ser Ser Asn 50 55 60

Pro Gly Thr Gln Val Tyr Glu Asp Tyr Asn Cys Thr Leu Asn Gln Thr 65 70 75 80

Asn Ile Glu Asn Asn Asn Lys Phe Tyr Ile Ile Gln Leu Leu Gln

				85					90					95	
Asp	Ser	Asn	Arg 100	Phe	Phe	Thr	Cys	Trp 105	Asn	Arg	Trp	Gly	Arg 110	Val	Gly
Glu	Val	Gly 115	Gln	Ser	Lys	Ile	Asn 120	His	Phe	Thr	Arg	Leu 125	Glu	Asp	Ala
Lys	Lys 130	Asp	Phe	Glu	Lys	Lys 135	Phe	Arg	Glu	Lys	Thr 140	Lys	Asn	Asn	Trp
Ala 145	Glu	Arg	Asp	His	Phe 150	Val	Ser	His	Pro	Gly 155	Lys	Tyr	Thr	Leu	Ile 160
Glu	Val	Gln	Ala	Glu 165	Asp	Glu	Ala	Gln	Glu 170	Ala	Val	Val	Lys	Val 175	Asp
Arg	Gly	Pro	Val 180	Arg	Thr	Val	Thr	Lys 185	Arg	Val	Gln	Pro	Cys 190	Ser	Leu
Asp	Pro	Ala 195	Thr	Gln	Lys	Leu	Ile 200	Thr	Asn	Ile	Phe	Ser 205	Lys	Glu	Met
Phe	Lys 210	Asn	Thr	Met	Ala	Leu 215	Met	Asp	Leu	Asp	Val 220	Lys	Lys	Met	Pro
Leu 225	Gly	Lys	Leu	Ser	Lys 230	Gln	Gln	Ile	Ala	Arg 235	Gly	Phe	Glu	Ala	Leu 240
Glu	Ala	Leu	Glu	Glu 245	Ala	Leu	Lys	Gly	Pro 250	Thr	Asp	Gly	Gly	Gln 255	Ser
Leu	Glu	Glu	Leu 260	Ser	Ser	His	Phe	Tyr 265	Thr	Val	Ile	Pro	His 270	Asn	Phe
Gly	His	Ser 275	Gln	Pro	Pro	Pro	Ile 280	Asn	Ser	Pro	Glu	Leu 285	Leu	Gln	Ala
Lys	Lys 290	Asp	Met	Leu	Leu	Val 295	Leu	Ala	Asp	Ile	Glu 300	Leu	Ala	Gln	Ala
Leu 305	Gln	Ala	Val	Ser	Glu 310	Gln	Glu	Lys	Thr	Val 315	Glu	Glu	Val	Pro	His 320
Pro	Leu	Asp	Arg	Asp 325	Tyr	Gln	Leu	Leu	Lys 330	Cys	Gln	Leu	Gln	Leu 335	Leu
Asp	Ser	Gly	Ala 340	Pro	Glu	Tyr	Lys	Val 345	Ile	Gln	Thr	Tyr	Leu 350	Glu	Gln
Thr	Gly	Ser 355	Asn	His	Arg	Cys	Pro 360	Thr	Leu	Gln	His	11e 365	Trp	Lys	Val
Asn	Gln 370	Glu	Gly	Glu	Glu	Asp 375	Arg	Phe	Gln	Ala	His 380	Ser	Lys	Leu	Gly
Asn 385	Arg	Lys	Leu	Leu	Trp 390	His	Gly	Thr	Asn	Met 395		Val	Val	Ala	Ala 400
Ile	Leu	Thr	Ser	Glv	Leu	Arq	Ile	Met	Pro	His	Ser	Gly	Gly	Arg	Val

405

СТА	гуѕ	GTÅ	420	ıyı	FIIC	AIG	261	425	ASII	Ser	шуз	Jer	430	GLY	TYL	
Val	Ile	Gly 435	Met	Lys	Cys	Gly	Ala 440	His	His	Val	Gly	Tyr 445	Met	Phe	Leu	
Gly	Glu 450	Val	Ala	Leu	Gly	Arg 455	Glu	His	His	Ile	Asn 460	Thr	Asp	Asn	Pro	
Ser 465	Leu	Lys	Ser	Pro	Pro 470	Pro	Gly	Phe	Asp	Ser 475	Val	Ile	Ala	Arg	Gly 480	
His	Thr	Glu	Pro	Asp 485	Pro	Thr	Gln	Asp	Thr 490	Glu	Leu	Glu	Leu	Asp 495	Gly	
Gln	Gln	Val	Val 500	Val	Pro	Gln	Gly	Gln 505	Pro	Val	Pro	Cys	Pro 510	Glu	Phe	
Ser	Ser	Ser 515	Thr	Phe	Ser	Gln	Ser 520	Glu	Tyr	Leu	Ile	Tyr 525	Gln	Glu	Ser	
Gln	Cys 530	Arg	Leu	Arg	Tyr	Leu 535	Leu	Glu	Val	His	Leu 540	*				
(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	10.	7:								
		(1	A) L1 3) T1 C) S1	ENGTI YPE: TRANI OPOLO	nuci nuci DEDNI DGY:	740 l leic ESS: lin	acio sino ear	pai: d	rs							
	•) MOI					A									
	(iii) HY	POTH	ETIC.	AL: 1	NO										
	(iv) AN'	ri s	ENSE	: NO											
	(vi) OR:					mus	culu	s							
	(ix		A) N.	E: AME/ OCAT				0								
	(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0: 7	:					
CCC	GGCT'	TTC .	ACTT	TTTC	TG C	TGCC	TCGG	G GA	ACAC	CTCG	AGC	CAAC	TGC	TTCC	TAACTC	60
AGG	GTGG	GCA (GAAC'	TGAC	GG G.	ATCT	AAGC	т тс	TGCA	тстс	TGA	GGAG	AAC		G GCT t Ala	117
							Gln					Lys			CGA Arg	165

CAA Gln 560	GGG Gly	ACA Thr	GAG Glu	GAG Glu	GAG Glu 565	GAC Asp	AGC Ser	TTC Phe	CGG Arg	TCC Ser 570	ACT Thr	GCC Ala	GAG Glu	GCT Ala	CTC Leu 575	213
AGA Arg	GCA Ala	GCA Ala	CCT Pro	GCT Ala 580	GAT Asp	AAT Asn	CGG Arg	GTC Val	ATC Ile 585	CGT Arg	GTG Val	GAC Asp	CCC Pro	TCA Ser 590	TGT Cys	261
CCA Pro	TTC Phe	AGC Ser	CGG Arg 595	AAC Asn	CCC Pro	GGG Gly	ATA Ile	CAG Gln 600	GTC Val	CAC His	GAG Glu	GAC Asp	TAT Tyr 605	GAC Asp	TGT Cys	309
												AAG Lys 620				357
												TGG Trp				405
												CAC His				453
CTG Leu	GAA Glu	GAT Asp	GCA Ala	AAG Lys 660	AAG Lys	GAC Asp	TTT Phe	AAG Lys	AAG Lys 665	AAA Lys	TTT Phe	TGG Trp	GAG Glu	AAG Lys 670	ACT Thr	501
AAA Lys	AAC Asn	AAA Lys	TGG Trp 675	GAG Glu	GAG Glu	CGG Arg	GAC Asp	CGT Arg 680	TTT Phe	GTG Val	GCC Ala	CAG Gln	CCC Pro 685	AAC Asn	AAG Lys	549
TAC Tyr	ACA Thr	CTT Leu 690	ATA Ile	GAA Glu	GTC Val	CAG Gln	GGA Gly 695	GAA Glu	GCA Ala	GAG Glu	AGC Ser	CAA Gln 700	GAG Glu	GCT Ala	GTA Val	597
GTG Val	AAG Lys 705	GCC Ala	TTA Leu	TCT Ser	CCC Pro	CAG Gln 710	GTG Val	GAC Asp	AGC Ser	GGC Gly	CCT Pro 715	GTG Val	AGG Arg	ACC Thr	GTG Val	645
						Asp						CTT Leu				693
ATC Ile	TTC Phe	AGC Ser	AAA Lys	GAG Glu 740	Met	TTC Phe	AAG Lys	AAC Asn	GCA Ala 745	Met	ACC Thr	CTC Leu	ATG Met	AAC Asn 750	CTG Leu	741
GAT Asp	GTG Val	AAG Lys	AAG Lys 755	Met	CCC Pro	TTG Leu	GGA Gly	AAG Lys 760	Leu	ACC Thr	AAG Lys	CAG Gln	CAG Gln 765	Ile	GCC Ala	789
CGT	GGC	TTC Phe 770	Glu	GCC Ala	TTG Leu	GAA Glu	GCT Ala 775	Leu	GAG Glu	GAG Glu	GCC Ala	ATG Met 780	Lys	AAC Asn	CCC Pro	837
ACA Thr	GGG Gly	GAT Asp	GGC Gly	CAG Gln	AGC Ser	CTG	GAA Glu	GAG Glu	CTC Leu	TCC Ser	TCC Ser	TGC Cys	TTC Phe	TAC	ACT Thr	885

	785					790					795					
			CAC His													933
			CTT Leu													981
			GCG Ala 835													1029
			GAG Glu													1077
			CTT Leu													1125
			TAC Tyr													1173
			GTT Val													1221
			TCC Ser 915													1269
			GTG Val													1317
			GGT Gly													1365
			TCA Ser													1413
			TAC Tyr													1461
			ATC Ile 995						Lys					Gly		1509
			Ile					Thr					Ala		GAC Asp	1557
ATT	GAA	CTT	GAA	CTG	GAT	GGG	CAG	CCG	GTG	GTG	GTG	ccc	CAA	GGC	CCG	1605

Ile	Glu 1025	Glu	Leu	Asp	Gly 1030		Pro	Val	Val	Val 1035	Gln	Gly	Pro		
	Val			TCA Ser 1045	Phe					Phe				165	3
				GAG Glu O					Leu				Glu	170	1
ATT Ile		 TAAG	GCTG	CTT (GCCC:	rccc:	ra G	GTCC	AAGC	2				174	0

(2) INFORMATION FOR SEQ ID NO: 8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 533 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Ala Pro Lys Arg Lys Ala Ser Val Gln Thr Glu Gly Ser Lys Lys
1 5 10 15

Gln Arg Gln Gly Thr Glu Glu Glu Asp Ser Phe Arg Ser Thr Ala Glu 20 25 30

Ala Leu Arg Ala Ala Pro Ala Asp Asn Arg Val Ile Arg Val Asp Pro 35 40 45

Ser Cys Pro Phe Ser Arg Asn Pro Gly Ile Gln Val His Glu Asp Tyr 50 55 60

Asp Cys Thr Leu Asn Gln Thr Asn Ile Gly Asn Asn Asn Asn Lys Phe 65 70 75 80

Tyr Ile Ile Gln Leu Leu Glu Glu Gly Ser Arg Phe Phe Cys Trp Asn 85 90 95

Arg Trp Gly Arg Val Gly Glu Val Gly Gln Ser Lys Met Asn His Phe 100 105 110

Thr Cys Leu Glu Asp Ala Lys Lys Asp Phe Lys Lys Phe Trp Glu

Lys Thr Lys Asn Lys Trp Glu Glu Arg Asp Arg Phe Val Ala Gln Pro 130 135 140

Asn Lys Tyr Thr Leu Ile Glu Val Gln Gly Glu Ala Glu Ser Gln Glu 145 150 155 160

Ala Val Val Lys Ala Leu Ser Pro Gln Val Asp Ser Gly Pro Val Arg 165 170 175

Thr Val Val Lys Pro Cys Ser Leu Asp Pro Ala Thr Gln Asn Leu Ile

180 185 190 Thr Asn Ile Phe Ser Lys Glu Met Phe Lys Asn Ala Met Thr Leu Met 200 Asn Leu Asp Val Lys Lys Met Pro Leu Gly Lys Leu Thr Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu Ala Met Lys 230 Asn Pro Thr Gly Asp Gly Gln Ser Leu Glu Glu Leu Ser Ser Cys Phe Tyr Thr Val Ile Pro His Asn Phe Gly Arg Ser Arg Pro Pro Pro Ile 265 Asn Ser Pro Asp Val Leu Gln Ala Lys Lys Asp Met Leu Leu Val Leu 280 Ala Asp Ile Glu Leu Ala Gln Thr Leu Gln Ala Ala Pro Gly Glu Glu Glu Glu Lys Val Glu Glu Val Pro His Pro Leu Asp Arg Asp Tyr Gln 315 310 Leu Leu Arg Cys Gln Leu Gln Leu Leu Asp Ser Gly Glu Ser Glu Tyr Lys Ala Ile Gln Thr Tyr Leu Lys Gln Thr Gly Asn Ser Tyr Arg Cys Pro Asn Leu Arg His Val Trp Lys Val Asn Arg Glu Gly Glu Gly Asp 360 Arg Phe Gln Ala His Ser Lys Leu Gly Asn Arg Arg Leu Leu Trp His 375 Gly Thr Asn Val Ala Val Val Ala Ala Ile Leu Thr Ser Gly Leu Arg Ile Met Pro His Ser Gly Gly Arg Val Gly Lys Gly Ile Tyr Phe Ala Ser Glu Asn Ser Lys Ser Ala Gly Tyr Val Thr Thr Met His Cys Gly Gly His Gln Val Gly Tyr Met Phe Leu Gly Glu Val Ala Leu Gly Lys 440 Glu His His Ile Thr Ile Asp Asp Pro Ser Leu Lys Ser Pro Pro Pro Gly Phe Asp Ser Val Ile Ala Arg Gly Gln Thr Glu Pro Asp Pro Ala Gln Asp Ile Glu Leu Glu Leu Asp Gly Gln Pro Val Val Val Pro Gln Gly Pro Pro Val Gln Cys Pro Ser Phe Lys Ser Ser Ser Phe Ser Gln

			500					505					510			
Ser	Glu	Tyr 515	Leu	Ile	Tyr	Lys	Glu 520	Ser	Gln	Cys	Arg	Leu 525	Arg	Tyr	Leu	
Leu	Glu 530	Ile	His	Leu												
(2)	INFC	RMAT	ON	FOR	SEQ	ID N	10: 9) :								
	(i)	(A (E (C	OUENC L) LE B) TY C) ST D) TO	NGTE PE:	nucl	87 b eic SS:	ase acio sino	pai:	:s							
	(ii)	MOI	ECUI	E TY	PE:	CDNA	1									
((iii)	нув	OTHE	TICA	AL: N	Ю										
	(iv)	ANT	ri se	NSE:	NO.											
	(vi)		GINA													
		(F	A) - OF	RGANI	SM:	Mus	muso	culus	5							
	(ix)	(P	ATURE A) NE B) LO	ME/F			584									
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTIC	ON: S	SEQ :	ID NO): 9:	:					
												GGC Gly				48
												TCC Ser				96
	CTC	A C A	CCA	GCA		CCT	САТ	ልልጥ	cee	GTC	ልጥሮ	CGT	GTG	GAC	CCC	144
												Arg				
												CAC				192
Ser	Cys	Pro	585	Ser	Arg	Asn	Pro	590	iie	GIN	vaı	His	595	ASP	TYL	
												AAC Asn 610				240
															AAT Asn	288
CGC	TGG	GGC	CGC	GTG	GGA	GAG	GTG	GGC	CAG	AGC	AAG	ATG	AAC	CAC	TTC	336

Arg Trp Gly Arg Val Gly Glu Val Gly Gln Ser Lys Met Asn His Phe

												AAA Lys				384	4
												GTG Val				43:	2
												GAG Glu 690				48	0
												GTG Val				52	8
TGC Cys 710	TCC Ser	CTA Leu	GAC Asp	CCT Pro	GCC Ala 715	ACC Thr	CAG Gln	AAC Asn	CTT Leu	ATC Ile 720	ACC Thr	AAC Asn	ATC Ile	TTC Phe	AGC Ser 725	57	6
												CTG Leu				62	4
AAG Lys	ATG Met	CCC Pro	TTG Leu 745	GGA Gly	AAG Lys	CTG Leu	ACC Thr	AAG Lys 750	CAG Gln	CAG Gln	ATT Ile	GCC Ala	CGT Arg 755	GGC Gly	TTC Phe	67	2
GAG Glu	GCC Ala	TTG Leu 760	GAA Glu	GCT Ala	CTA Leu	GAG Glu	GAG Glu 765	GCC Ala	ATG Met	AAA Lys	AAC Asn	CCC Pro 770	ACA Thr	GGG Gly	GAT Asp	72	0
		Ser										ACT Thr				76	8
	Asn											TCC Ser				81	.6
CTT Leu	CAG Gln	GCC Ala	AAG Lys	AAG Lys 810	Asp	ATG Met	CTG Leu	CTG Leu	GTG Val 815	CTA Leu	GCG Ala	GAC Asp	ATC Ile	GAG Glu 820	TTG Leu	86	4
									Glu			GAG Glu		Val	GAA Glu	91	. 2
												CTC Leu 850	Arg		CAG Gln	96	50
		Leu					Glu					Ala			ACC Thr	100	8(
												AAC Asn			CAT	105	56

870					875					880					885	
GTT Val	TGG Trp	AAA Lys	GTG Val	AAC Asn 890	CGA Arg	GAA Glu	GGG Gly	GAG Glu	GGA Gly 895	GAC Asp	AGG Arg	TTC Phe	CAG Gln	GCC Ala 900	CAC His	1104
			GGC Gly 905													1152
			GCC Ala													1200
GGT Gly	GGT Gly 935	CGT Arg	GTT Val	GGC Gly	AAG Lys	GGT Gly 940	ATT Ile	TAT Tyr	TTT Phe	GCC Ala	TCT Ser 945	GAG Glu	AAC Asn	AGC Ser	AAG Lys	1248
TCA Ser 950	GCT Ala	GGC Gly	TAT Tyr	GTT Val	ACC Thr 955	ACC Thr	ATG Met	CAC His	TGT Cys	GGG Gly 960	GGC Gly	CAC His	CAG Gln	GTG Val	GGC Gly 965	1296
TAC Tyr	ATG Met	TTC Phe	CTG Leu	GGC Gly 970	GAG Glu	GTG Val	GCC Ala	CTC Leu	GGC Gly 975	AAA Lys	GAG Glu	CAC His	CAC His	ATC Ile 980	ACC Thr	1344
ATC Ile	GAT Asp	GAC Asp	CCC Pro 985	AGC Ser	TTG Leu	AAG Lys	AGT Ser	CCA Pro 990	CCC Pro	CCT Pro	GGC Gly	TTT Phe	GAC Asp 995	AGC Ser	GTC Val	1392
ATC Ile	GCC Ala	CGA Arg	GGC Gly 0	CAA Gln	ACC Thr	GAG Glu	CCG Pro	Asp	CCC Pro	GCC Ala	CAG Gln	GAC Asp	Ile	GAA Glu	CTT Leu	1440
GAA Glu	CTG Leu 101	Asp	GGG Gly	CAG Gln	CCG Pro	GTG Val 102	Val	GTG Val	CCC Pro	CAA Gln	GGC Gly 102	Pro	CCT Pro	GTG Val	CAG Gln	1488
TGC Cys	Pro	TCA Ser	TTC Phe	AAA Lys	AGC Ser 103	Ser	AGC Ser	TTC Phe	AGC Ser	CAG Gln 104	Ser	GAA Glu	TAC Tyr	CTC Leu	ATA Ile 1045	1536
TAC Tyr	AAG Lys	GAG Glu	AGC Ser	CAG Gln 105	Cys	CGC Arg	CTG Leu	CGC Arg	TAC Tyr 105	Leu	CTG Leu	GAG Glu	ATT Ile	CAC His 106	Leu	1584
TAA																1587

(2) INFORMATION FOR SEQ ID NO: 10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 528 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Ala Pro Lys Arg Lys Ala Ser Val Gln Thr Glu Gly Ser Lys Lys

1				5					10					15	
Gln	Arg	Gln	Gly 20	Thr	Glu	Glu	Glu	Asp 25	Ser	Phe	Arg	Ser	Thr 30	Ala	Glu
Ala	Leu	Arg 35	Ala	Ala	Pro	Ala	Asp 40	Asn	Arg	Val	Ile	Arg 45	Val	Asp	Pro
Ser	Cys 50	Pro	Phe	Ser	Arg	Asn 55	Pro	Gly	Ile	Gln	Val 60	His	Glu	Asp	Tyr
Asp 65	Cys	Thr	Leu	Asn	Gln 70	Thr	Asn	Ile	Gly	Asn 75	Asn	Asn	Asn	Lys	Phe 80
Tyr	Ile	Ile	Gln	Leu 85	Leu	Glu	Glu	Gly	Ser 90	Arg	Phe	Phe	Cys	Trp 95	Asn
Arg	Trp	Gly	Arg 100	Val	Gly	Glu	Val	Gly 105	Gln	Ser	Lys	Met	Asn 110	His	Phe
Thr	Cys	Leu 115	Glu	Asp	Ala	Lys	Lys 120	Asp	Phe	Lys	Lys	Lys 125	Phe	Trp	Glu
_	130	_			Trp	135					140				
Asn 145	Lys	Tyr	Thr	Leu	Ile 150	Glu	Val	Gln	Gly	Glu 155	Ala	Glu	Ser	Gln	Glu 160
Ala	Val	Val	Lys	Val 165	Asp	Ser	Gly	Pro	Val 170	Arg	Thr	Val	Val	Lys 175	Pro
Cys	Ser	Leu	Asp 180	Pro	Ala	Thr	Gln	Asn 185	Leu	Ile	Thr	Asn	Ile 190	Phe	Ser
Lys	Glu	Met 195	Phe	Lys	Asn	Ala	Met 200	Thr	Leu	Met	Asn	Leu 205	Asp	Val	Lys
Lys	Met 210	Pro	Leu	Gly	Lys	Leu 215	Thr	Lys	Gln	Gln	Ile 220	Ala	Arg	Gly	Phe
Glu 225	Ala	Leu	Glu	Ala	Leu 230	Glu	Glu	Ala	Met	Lys 235	Asn	Pro	Thr	Gly	Asp 240
Gly	Gln	Ser	Leu	Glu 245	Glu	Leu	Ser	Ser	Cys 250	Phe	Tyr	Thr	Val	Ile 255	Pro
His	Asn	Phe	Gly 260	Arg	Ser	Arg	Pro	Pro 265		Ile	Asn	Ser	Pro 270	Asp	Val
Leu	Gln	Ala 275	Lys	Lys	Asp	Met	Leu 280	Leu	Val	Leu	Ala	Asp 285		Glu	Leu
Ala	Gln 290	Thr	Leu	Gln	Ala	Ala 295		Gly	Glu	Glu	Glu 300		Lys	Val	Glu
Glu 305		Pro	His	Pro	Leu 310		Arg	Asp	Tyr	Gln 315		Leu	Arg	Cys	Gln 320
Leu	Gln	Leu	Leu	Asp	Ser	Glv	Glu	Ser	Glu	Tyr	Lys	Ala	Ile	Gln	Thr

330 325 335 Tyr Leu Lys Gln Thr Gly Asn Ser Tyr Arg Cys Pro Asn Leu Arg His Val Trp Lys Val Asn Arg Glu Gly Glu Gly Asp Arg Phe Gln Ala His 360 Ser Lys Leu Gly Asn Arg Arg Leu Leu Trp His Gly Thr Asn Val Ala 375 Val Val Ala Ala Ile Leu Thr Ser Gly Leu Arg Ile Met Pro His Ser Gly Gly Arg Val Gly Lys Gly Ile Tyr Phe Ala Ser Glu Asn Ser Lys 410 Ser Ala Gly Tyr Val Thr Thr Met His Cys Gly Gly His Gln Val Gly 420 Tyr Met Phe Leu Gly Glu Val Ala Leu Gly Lys Glu His His Ile Thr 440 Ile Asp Asp Pro Ser Leu Lys Ser Pro Pro Pro Gly Phe Asp Ser Val Ile Ala Arg Gly Gln Thr Glu Pro Asp Pro Ala Gln Asp Ile Glu Leu 470 475 Glu Leu Asp Gly Gln Pro Val Val Val Pro Gln Gly Pro Pro Val Gln Cys Pro Ser Phe Lys Ser Ser Ser Phe Ser Gln Ser Glu Tyr Leu Ile 505 Tyr Lys Glu Ser Gln Cys Arg Leu Arg Tyr Leu Leu Glu Ile His Leu

520

(2) INFORMATION FOR SEQ ID NO: 11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: YES
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:2
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer 1 bis 5 andere Aminosaeuren"
- (ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION: 3
- (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

Pro Xaa Xaa Gly Xaa Xaa Gly Lys Gly Ile Tyr Phe Ala 1 5 10

- (2) INFORMATION FOR SEQ ID NO: 12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:1
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:6
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ile oder Val"
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:9
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer 1 bis 5 andere Aminosaeuren"
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:10
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Xaa Xaa Gly Leu Arg Xaa Xaa Pro Xaa Xaa Gly Xaa Xaa Xaa Gly Lys
1 10 15

Gly Ile Tyr Phe Ala 20

- (2) INFORMATION FOR SEQ ID NO: 13:
 - (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: YES
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:16
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:21
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ile oder Val"
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:24
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer 1 bis 5 andere Aminosaeuren"
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:25
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:6
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ser oder Thr"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

Leu Leu Trp His Gly Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ile Leu Xaa 1 5 10 15

Xaa Gly Leu Arg Xaa Xaa Pro Xaa Xaa Gly Xaa Xaa Xaa Gly Lys Gly
20 25 30

Ile Tyr Phe Ala Xaa Xaa Xaa Ser Lys Ser Ala Xaa Tyr 35 40 45

- (2) INFORMATION FOR SEQ ID NO: 14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: YES
- (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:1
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Leu oder Val"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

Xaa Xaa Xaa Xaa Xaa Xaa Leu Xaa Xaa Xaa Xaa Xaa Xaa Leu Xaa 1 10 15

Xaa Xaa Xaa Xaa Leu 20

- (2) INFORMATION FOR SEQ ID NO: 15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:21
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Asp oder Glu"
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION:22
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer 10 oder 11 andere Aminosaeuren"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

Leu Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Asn Xaa Xaa Tyr Xaa Xaa 1 5 10 15

Gln Leu Leu Xaa Xaa Trp Gly Arg Val Gly

- (2) INFORMATION FOR SEQ ID NO: 16:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 amino acids
 - (B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: YES
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

Ala Xaa Xaa Xaa Phe Xaa Lys Xaa Xaa Xaa Lys Thr Xaa As
n Xaa 1 5 10 15

Trp Xaa Xaa Xaa Xaa Aaa Phe Xaa Xaa Pro Xaa Lys
20 25

- (2) INFORMATION FOR SEQ ID NO: 17:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 44 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION: 4
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Ile oder Leu"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

Gln Xaa Leu Xaa Xaa Xaa Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa 1 10 15

Met Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Pro Leu Gly Lys Leu 20 25 30

 x_{aa} x_{\text

- (2) INFORMATION FOR SEQ ID NO: 18:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

Phe Tyr Thr Xaa Ile Pro His Xaa Phe Gly Xaa Xaa Xaa Pro Pro 1 5 10 15

- (2) INFORMATION FOR SEQ ID NO: 19:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

Lys Xaa Xaa Xaa Leu Xaa Xaa Leu Xaa Asp Ile Glu Xaa Ala Xaa Xaa $1 \hspace{1.5cm} 5 \hspace{1.5cm} 10 \hspace{1.5cm} 15$

Leu

- (2) INFORMATION FOR SEQ ID NO: 20:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 11 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

Gly Xaa Xaa Xaa Leu Xaa Glu Val Ala Leu Gly
1 5 10

- (2) INFORMATION FOR SEQ ID NO: 21:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (ix) FEATURE:

- (A) NAME/KEY: Region
- (B) LOCATION:14
- (D) OTHER INFORMATION:/note= "Xaa steht fuer 7 bis 9 andere Aminosaeuren"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

Gly Xaa Xaa Ser Xaa Xaa Xaa Gly Xaa Xaa Pro Xaa Leu Xaa 1 5 10 15

Gly Xaa Xaa Val 20

- (2) INFORMATION FOR SEQ ID NO: 22:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: YES
 - (ix) FEATURE:
 - (A) NAME/KEY: Region
 - (B) LOCATION: 2
 - (D) OTHER INFORMATION:/note= "Xaa steht fuer Tyr oder Phe"
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

Glu Xaa Xaa Xaa Tyr Xaa Xaa Xaa Gln Xaa Xaa Xaa Xaa Tyr Leu Leu 1 5 10 15

- (2) INFORMATION FOR SEQ ID NO: 23:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

Leu Asn Glu Ser

- (2) INFORMATION FOR SEQ ID NO: 24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

Lys Thr Glu Leu Gln Ser Pro Glu His Pro Leu Asp Gln His Tyr Arg
1 10 15

Asn Leu His Cys 20

- (2) INFORMATION FOR SEQ ID NO: 25:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

Ala Glu Ala Leu Lys 20

- (2) INFORMATION FOR SEQ ID NO: 26:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

0050/49100/49790

Cys Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu 1 5 10 15

Glu Ala Leu Lys 20

- (2) INFORMATION FOR SEQ ID NO: 27:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 19 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu 1 5 10 15

Ala Leu Lys

- (2) INFORMATION FOR SEQ ID NO: 28:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 19 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu 1 5 10 15

Ala Met Lys

coroasea.a*M*W

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09/70**1**586 **529 Rec**'d PCT/PTC **3**0 NOV 2000

We claim:

- A poly(ADP-ribose) polymerase (PARP) homolog derived from a human or non-human mammal which has an amino acid sequence which has
 - a) a functional NAD+ binding domain
 and
 - b) no zinc finger sequence motif of the general formula

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 $CX_2CX_mHX_2C$

in which

m is an integral value from 28 or 30, and the X radicals are, independently of one another, any amino acid.

2. A PARP homolog as claimed in claim 1, wherein the functional NAD+ binding domain comprises one of the following general sequence motifs:

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$$\label{eq:continuous} \begin{split} PX_n(S/T)GX_3GKGIYFA,\\ (S/T)XGLR(I/V)XPX_n(S/T)GX_3GKGIYFA \text{ or }\\ LLWHG(S/T)X_7IL(S/T)XGLR(I/V)XPX_n(S/T)GX_3GKGIYFAX_3SKSAXY \end{split}$$

25 in which

n is an integral value from 1 to 5, and the X radicals are, independently of one another, any amino acid.

3. A PARP homolog as claimed in either of the preceding claims, 30 comprising at least another one of the following part-sequence motifs:

> LX₉NX₂YX₂QLLX(D/E)X_{10/11}WGRVG, AX₃FXKX₄KTXNXWX₅FX₃PXK, QXL(I/L)X₂IX₉MX₁₀PLGKLX₃QIX₆L, FYTXIPHXFGX₃PP; and KX₃LX₂LXDIEXAX₂L,

in which the X radicals are, independently of one another,
any amino acid.

4. A PARP homolog as claimed in any of the preceding claims, selected from human PARP homologs, which has the amino acid sequence shown in SEQ ID NO: 2 (human PARP2) or SEQ ID NO: 4 or 6 (human PARP3 type 1 or 2); or murine PARP homologs which

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have the amino acid sequence shown in SEQ ID NO:8 (mouse PARP long form) or SEQ ID No:10 (mouse PARP short form).

- 5. A binding partner having specifity for PARP homologs as claimed in any of the preceding claims, selected from
 - a) antibodies and fragments thereof,
 - b) protein-like compounds which interact with a part-sequence of the protein, and
- c) low molecular weight effectors which modulate the catalytic PARP activity or another biological function of a PARP molecule.
 - 6. A nucleic acid comprising
- a) a nucleotide sequence coding for at least one PARP homolog as claimed in any of claims 1 to 4, or the complementary nucleotide sequence thereof;
 - b) a nucleotide sequence which hybridizes with a sequence as specified in a) under stringent conditions; or
- c) nucleotide sequences which are derived from the nucleotide sequences defined in a) and b) through the degeneracy of the genetic code.
 - 7. A nucleic acid as claimed in claim 6, comprising
- 25 a) nucleotides +3 to +1715 shown in SEQ ID NO:1;
 - b) nucleotides +242 to +1843 shown in SEQ ID NO:3;
 - c) nucleotides +221 to +1843 shown in SEQ ID NO:5;
 - d) nucleotides +112 to +1710 shown in SEQ ID NO:7; or
 - e) nucleotides +1 to +1584 shown in SEQ ID NO:9.

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- 8. An expression cassette comprising, under the genetic control of at least one regulatory nucleotide sequence, at least one nucleotide sequence as claimed in either of claims 6 and 7.
- 35 9. A recombinant vector comprising at least one expression cassette as claimed in claim 8.
 - 10. A recombinant microorganism comprising at least one recombinant vector as claimed in claim 9.

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11. A transgenic mammal comprising a vector as claimed in claim 9.

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- 12. A PARP-deficient mammal or PARP-deficient eukaryotic cell, in which functional expression of at least one gene which codes for a PARP homolog as claimed in any of claims 1 to 4 is inhibited.
- 13. An in vitro detection method for PARP inhibitors, which comprises
- a) incubating an unsupported or supported polyADP-ribosylatable target with a reaction mixture comprising
 - al) a PARP homolog as claimed in any of claims 1 to 4,
 - a2) a PARP activator; and
 - a3) a PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected;
 - b) carrying out the polyADP ribosylation reaction; and
 - c) determining the polyADP ribosylation of the target qualitatively or quantitatively.
- 20 14. A method as claimed in claim 13, wherein the PARP homolog is preincubated with the PARP activator and the PARP inhibitor or an analyte in which at least one PARP inhibitor is suspected, before the polyADP ribosylation reaction is carried out.
 - 15. A method as claimed in either of claims 13 and 14, wherein the polyADP-ribosylatable target is a histone protein.
- 16. A method as claimed in any of claims 13 to 15, wherein the PARP activator is activated DNA.
 - 17. A method as claimed in any of claims 13 to 16, wherein the polyADP ribosylation reaction is started by adding NAD+.
- 35 18. A method as claimed in any of claims 13 to 17, wherein the polyADP ribosylation of the supported target is determined using anti-poly(ADP-ribose) antibodies.
- 19. A method as claimed in any of claims 13 to 17, wherein the unsupported target is labeled with an acceptor fluorophore.
- 20. A method as claimed in claim 19, wherein the polyADP ribosylation of the unsupported target is determined using anti-poly(ADP-ribose) antibody which is labeled with a donor fluorophore which is able to transfer energy to the acceptor fluorophore.

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- 21. A method as claimed in either of claims 19 and 20, wherein the target is biotinylated histone, and the acceptor fluorophore is coupled thereto via avidin or streptavidin.
- 5 22. A method as claimed in either of claims 20 and 21, wherein the anti-poly(ADP-ribose) antibody carries a europium cryptate as donor fluorophore.
- 23. An in vitro screening method for binding partners for a PARP10 molecule, which comprises
 - al) immobilizing at least one PARP homolog as claimed in any of claims 1 to 4 on a support;
 - b1) contacting the immobilized PARP homolog with an analyte in which at least one binding partner is suspected; and
- c1) determining, where appropriate after an incubation period, analyte constituents bound to the immobilized PARP homolog;

or

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- a2) immobilizing on a support an analyte which comprises at least one possible binding partner for a PARP molecule;
- b2) contacting the immobilized analyte with at least one PARP homolog as claimed in any of claims 1 to 4 for which a binding partner is sought; and
- c2) examining the immobilized analyte, where appropriate after an incubation period, for binding of the PARP homolog.
- 30 24. A method for the qualitative or quantitative determination of nucleic acids encoding a PARP homolog as claimed in any of claims 1 to 4, which comprises
- a) incubating a biological sample with a defined amount of an exogenous nucleic acid as claimed in either of claims
 35 6 and 7, hybridizing under stringent conditions, determining the hybridizing nucleic acids and, where appropriate, comparing with a standard; or
- b) incubating a biological sample with a pair of
 oligonucleotide primers with specificity for a PARP
 homolog-encoding nucleic acid, amplifying the nucleic
 acid, determining the amplification product and, where
 appropriate, comparing with a standard.
- 45 25. A method for the qualitative or quantitative determination of a PARP homolog as claimed in any of claims 1 to 4, which comprises

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- a) incubating a biological sample with a binding partner specific for a PARP homolog,
- detecting the binding partner/PARP complex and, where appropriate,
- 5 c) comparing the result with a standard.
 - 26. A method as claimed in claim 25, wherein the binding partner is an antibody or a binding fragment thereof, which carries a detectable label where appropriate.

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- 27. A method as claimed in any of claims 24 to 26 for diagnosing energy deficit-mediated illnesses.
- 28. A method for determining the efficacy of PARP effectors,which comprises
 - a) incubating a PARP homolog as claimed in any of claims 1 to 4 with an analyte which comprises an effector of a physiological or pathological PARP activity; removing the effector again where appropriate; and
- 20 b) determining the activity of the PARP homolog, where appropriate after adding substrates or cosubstrates.
 - 29. A gene therapy composition, which comprises in a vehicle acceptable for gene therapy a nucleic acid construct which
- 25 a) comprises an antisense nucleic acid against a coding nucleic acid as claimed in either of claims 6 and 7; or
 - b) a ribozyme against a nucleic acid as claimed in either of claims 6 and 7; or
 - c) codes for a specific PARP inhibitor.

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- 30. A pharmaceutical composition comprising, in a pharmaceutically acceptable vehicle, at least one PARP protein as claimed in any of claims 1 to 4, at least one PARP binding partner as claimed in claim 5 or at least one coding nucleotide sequence as claimed in claim 6 or 7.
- 31. The use of low molecular weight PARP binding partners as claimed in claim 5 for the manufacture of a pharmaceutical agent for the diagnosis or therapy of pathological states in the development and/or progress of which at least one PARP protein, or a polypeptide derived therefrom, is involved.
- 32. The use of low molecular weight PARP binding partners as claimed in claim 5 for the manufacture of a pharmaceutical agent for the diagnosis or therapy of pathological states mediated by an energy deficit.

Abstract

The invention relates to poly(ADP-ribose)polymerase (PARP) 5 homologs which have an amino acid sequence which has

- a functional NAD+ binding domain and
- b) no zinc finger sequence motif of the general formula

 $CX_2CX_mHX_2C$

in which

m is an integral value from 28 or 30, and the X radicals are, independently of one another, any amino acid;

and the functional equivalents thereof; nucleic acids coding
15 therefor; antibodies with specificity for the novel protein;
pharmaceutical and gene therapy compositions which comprise
products according to the invention; methods for the analytical
determination of the proteins and nucleic acids according to the
invention; methods for identifying effectors or binding partners

20 of the proteins according to the invention; novel PARP effectors; and methods for determining the activity of such effectors.

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					1	Majority
5 2 3 2 2 2	70 G II S I R II P D V E V D G F S E L	80 R W D D Q Q K V K K T A	90 E A G G V T G K G Q T G G G R A	100 D G I G S K A E K T	110 T C O D F A A E Y A K S A L H E) ∫ human PARP J human PARP 2 human PARP 3 murine PARP
	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	3 4	K		170 180	Hajority J
121 20 3	N R S T C K G C M E K I E K G Q V	R C S K K H V D P E K P P W W W W W W W W W W W W W W W W W	SPAKTER	POCPVKNREE	LGPRPEYSA	humanPARP1 humanPARP2 humanPARP3 murinePARP
u	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	200 200 2	- D K - D	220	230 240	Majority
181	LXGFSLLATEDKEALKK	V K S E G K H K	X Y D Q V D B X O D X Y O	A X X X X X X X X X X X X X X X X X X X	K D S K L E K A L K A	humanPARP1 humanPARP2 humanPARP3 murinePARP
		X 1 1 - X -		1	1000 1000	Majority
1 4 1	250 Q N D L I W N I K D E L K K V C S '	260 T N D L K E L L I F N X E S V K A L L L K G K	0 0 0 V P S G E S A I	LDRVADGHVF	G A L L P C E E C S	humanPARP1 humanPARP2 humanPARP3 murinePARP
~			1(1)			-

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Majority 1 human PARP1 human PARP2 human PARP3 muxine PARP	Majority 0 1 humanPARP1 humanPARP2 murinePARP3	Hajority 0 ∫ humanPARP1 humanPARP2 humanPARP3 muzinePARP3	Majority) humanPARF1 humanPARP2 humanPARP3 murinePARP	Majority humanPARP1 humanPARP2 humanPARP3
360 K Q D R I F P P E	420 I E K L G G X L T	480 LFLAHILLSP	A V D P D S G L E	6 00 6 K
350 350 370 370 370 370 370 370 370 370 370 37	410 S.R.N.K.D.E.V.K.A.H.	470 470 470 470 470	M K L T L K G G A	Y W I F R S W O R V F T C W W R W G R V F T C W W R W G R V F T C W W R W G R V F T C W W R W G R V G R V F T C W W R W G R V F T C W W R W G R V G R V C W R W G R V C W C R V C W C R V
X W V	400 400 8 M K J L T L G K L	R S T A E A L	X E D 520 K E E G I N K S E K R Y E D	10 L L E D D X - R F 580 L Q L L E D D K E N R 10 L L E D D A Q R N 10 L L Q D S N - R P
M V K T Q T P N R K 130 M V K T Q T P N R K	390 390 S; S; A; S; A; D; K; P; L; S;	R Q X X X E B D X P 450 K M E E V K B A N I R Q A G R E E D P P F F R Q G T E E E D S F F	S10 S10 S10 S10 S10 S10 S10 S10	STO STO T V K G T N S Y Y K L Q P N N N K Y Y L I G N N N N K Y Y L
1 G D V T A W T K C	3 H O B O B O B O B O B O B O B O B O B O	T K E V E K M N K K K K K K K K K K K K K K K K K	S00 V V A P R G K S G A P K R I I R V D P T C B H R V J R V D P S C B	Y D C T L N Q T N S60 S60 F S A T L G L V D Y D V M L N Q T N Y D C T L N Q T N
M C L V F K S D A Y Y C K X V G K A H V Y C C	T S A S V A A T P P P P P P P P P P P P P P P P P	4 J O T A N K A S L C I S	490 W G A E V K A E P V E V	SS0
301 88 9 2	361 100 18	421 100 22 15	481 100 44 35	541 100 73

		3/7		
Majority) humanPARP1 humanPARP2 humanPARP3 murinePARP	Majority humanPARP1 humanPARP2 humanPARP3	Majority humanPAMP1 humanPAMP3 murinePAMP	Majority humanPARP1 humanPARP3 muranPARP3	Majority humanPARP1 humanPARP3 murinePARP
L N II F T X - L E D A K E D F X K K F X E K T K N N W E E R D X F V X X P G K Y T L L E V D Y - X E X E D E U A V V K - Ma 610 660 L E Q H P S K - L E D A I E II F H K L Y E B K T G N A W II S K N - F T K Y P K K P Y P L E I D Y G 0 D E E A V K X - hu L V A C S G H L H K A K D P E K K F R E K T K N N W A E R D H P V S H P G K Y T L I E V O A E D E A O E A V K A - hu I N H P T R - L E D A K K D P E K K F W E K T K N N W A E R D H P V S H P G K Y T L I E V O G E A E S Q E A V V K A - hu H N H F T C - L E D A K K D P K K F W E K T K W K W E E R D R F V A Q P N K Y T L I E V O G E A E S Q E A V K A - hu	- S L X V D X G P V S T V X K R V Q P C S L D P A T Q X L I T N I F S V E M P K N A M X L M X L D V K K M P L G K L S K Machine A C C C C C C C C C C C C C C C C C C	O Q 1 A A G F E A L E A L E E A X K X G T X G G Q S L E E L S S X P Y T V I P H D P G X S X P P L I N S P D X L Q A K X MAJ 730 780 R Q 1 Q A A Y S I L S E V Q Q A V S Q O S S D S Q 1 L D - L S N R F Y T L I P H D P G N K K P P L L N N A D S V Q A K V V N U P H N P G L S E K I H N U P G Q O 1 A R G F E A L E E A H K N P T G D G Q S L E E L S S C F Y T V I P H N P G R S R P P P I N S P D V L Q A K K MU P U Q Q I A R G F E A L E E A H K N P T G D G Q S L E E L S S C F Y T V I P H N F G R S R P P P I N S P D V L Q A K K MU P U Q Q I A R G F E A L E E A H K N P T G D G Q S L E E L S S C F Y T V I P H N F G R S R P P P I N S P D V L Q A K K MU P U D Q Q I A R G F E A L E E A L E E A L E A L E E A L E A L E E A H K N P T G D G Q S L E E L S S C F Y T V I P H N F G R S R P P P I N S P D V L Q A K K MU P U D Q Q I A R G F E A L E L E	DHLLVLADIELAGXCGAXXXEXSXXVEEVPHPLDRDYQLLXCQLQLLDSOSXEYXVIQTY Maj 790 800 810 820 840 840 EHLDNLLDIEVAYSLLRGGSDDSSKDPIDVNYEKLXTDIXVVDRDSEEAEIIRKY Num QLLEALDNLLDIEVAYSLLRGGSDDSSKHPLDQHYRNLHCALRPLDHESYEFKVISOY hum DMLLVLADIELAGOIELAGAYS-EQEKTVEEVPHPLDRDYQLLXCQLQLLDSORPEYXVIOTY hum DMLLVLADIELAGOIELAGAYS-EQEKTVEEVPHPLDRDYQLLXCQLQLDSORPEYXVIOTY mux	LKQTGAXTHCPYTLXDIPKVEREGEXDRFQAHSKLONRRLLWHGSNBAVVAGILSSGL Majorkatteren and statement of the statement of
601 149 119	655 209 175 166	704 260 231 223	763 319 291 283	818 373 350 343

Fig. 1(3

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L X X A Majority	LKIIA humanpARP1 LLE EA humanpARP2 INTD humanpARP3 ITID murinePARP	S L - Y S Majority 1020	T S L L Y N human PARP1 Y T L N Y N human PARP2 P S Q S human PARP3 P S Q S muxine PARP3	Majority	human PARP1 human PARP2 human PARP3 murine PARP
XEX	N O R X N O R N	S X P	5 5 0 V N D O O D T G I L N P D O O P C P E F S S S T O C P S F K S S S		
S X C G G H X V O L H L L D E V A L G 940 950	2) G D P 1 O L I R L K N T O L L L K C G A II H V O Y H II C G G II Q V G Y N	TEPDPAGDIELELDGGGVVVPLGPPVXCGXFXS	T T P D P S A N I S L D 0 V D V P L G T Q I H A P S S A H P V T L N 0 S T V P L G P A S T E P D P T Q D T E L E L D G Q Q V V P 0 G Q P V T E P D P A Q D I E L E L D G Q O V V P 0 G P P V		
5 X S A G Y V X T	Y S K S A N Y C H T S S S K S A N Y C P A S S N N Y C P A S S N N S K S A G Y V T T H	A Q D I E L E L D G	PSANISLD SAHPVTLN PTQDTELELDG	XXLW-	T S L W .
V G K G I Y P A S E N 920	F G K G I Y F A D V G K G I Y F A S V G K G I Y F A S	VIGLGKTEPDIV	TKGLGK TKGLGK VIARGH	Y L L E V H P N P - 3 1040	Y L L K L K F N F K T Y L L K V O P N P - L Y L L E V H L .
IAPHEAP-SGGR	A P P E A P V T G Y H A P P E A P I I T G Y H M P H S G G R M P H S G G R M P H S G G R M P M	PSLXSLPPGKDS	H I S K - L P K G K H S P K B C L L O G K H S P S L K S P P P G P D S P S L K S P P P G F D S	Y L V Y X E S Q V R L R	1 V Y D I A Q V N L K 1 V Y N P P N Q V R H R L I Y Q E S O C R L R L I Y K E S Q C R L R
2	878 R 431 R 407 R	z	916 S 489 N 463 N 456 D	ш	988 E Y 545 E Y 521 E Y 514 E Y

Fig. 1(4

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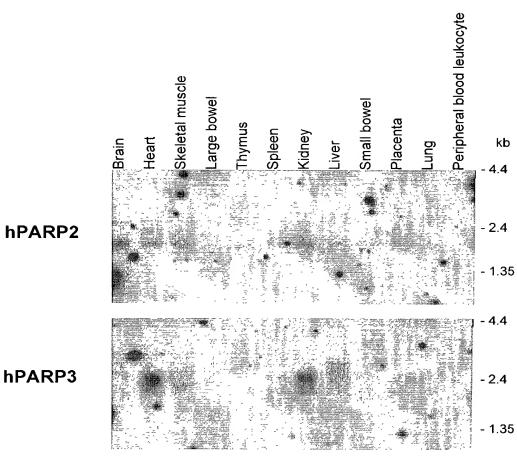


Fig. 2

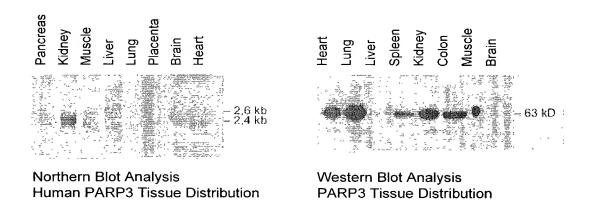


Fig. 3

Fig. 4

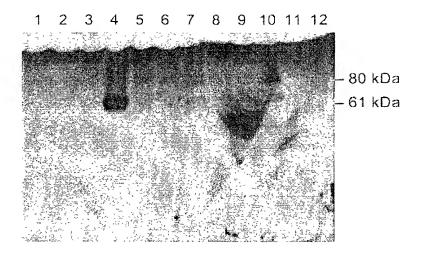
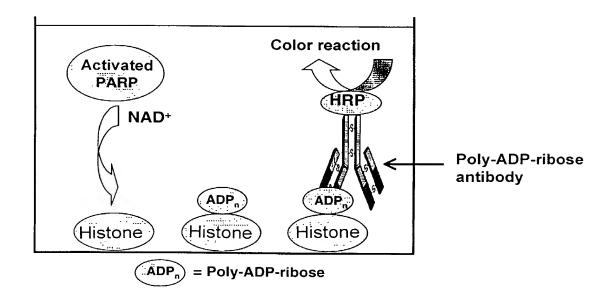
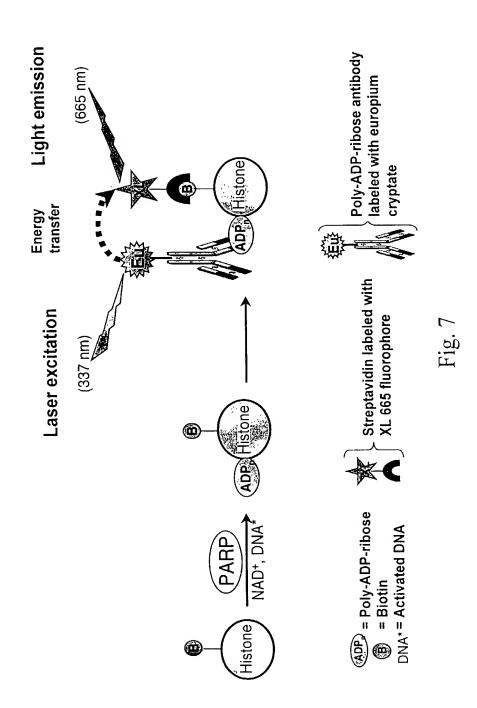


Fig. 5



HRP = Horseradish-Peroxidase

Fig. 6



Declaration, Power of Attorney

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0050/049100

We (I), the undersigned inventor(s), hereby declare(s) that:

My residence, post office address and citizenship are as stated below next to my name,

We (I) believe that we are (I am) the original, first, and joint (sole) inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled

Poly(ADP-ribose)polymerase-gene

the specification of which	
is attached hereto.	
[] was filed on	as
Application Serial No	
and amended on	•
[x] was filed as PCT interna	itional application
Number PCT/EP 99	9/ 03889
on <u>June 4, 1</u>	1999
and was amended under	PCT Article 19
	('C1'1 1 -)

- We (I) hereby state that we (I) have reviewed and understand the contents of the above—identified specification, including the claims, as amended by any amendment referred to above.
- We (I) acknowledge the duty to disclose information known to be material to the patentability of this application as defined in Section 1.56 of Title 37 Code of Federal Regulations.
- We (I) hereby claim foreign priority benefits under 35 U.S.C. § 119(a)—(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s)

Application No.	Country	Day/Month/Year	Priority Claimed
19825213.7	Germany	05 June 1998	[x] Yes [] No
19908837.3	Germany	01 March 1999	[x] Yes [] No

Declaration

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0050/049100

	Number)	(Filing Date)
(Application N	Number)	(Filing Date)
nternational application designating f this application is not disclosed in rst paragraph of 35 U.S.C. § 112, I a	g the United States, listed below a n the prior United States or PCT Ir acknowledge the duty to disclose ir	United States application(s), or § 365(c) of any P and, insofar as the subject matter of each of the claimternational application in the manner provided by information which is material to patentability as define prior application and the national or PCT Internation
Application Serial No.	Filing Date	Status (pending, patented, abandoned)

We (I) declare that all statements made herein of our (my) own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

And we (I) hereby appoint Messrs. HERBERT. B. KEIL, Registration Number 18,967; and RUSSEL E. WEINKAUF, Registration Number 18,495; the address of both being Messrs. Keil & Weinkauf, 1101 Connecticut Ave., N.W., Washington, D.C. 20036 (telephone 202–659–0100), our attorneys, with full power of substitution and revocation, to prosecute this application, to make alterations and amendments therein, to sign the drawings, to receive the patent, and to transact all

business in the Patent Office connected therewith.

nornysta la zono

Declaration Page 3 of 4 0050/049100

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0050/049100

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c'd PCT/PTO 25 APR 2002





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Asp Tyr Asn Cys Thr Leu Asn Gln Thr Asn Ile Glu Asn Asn Asn 65 70 75 80

Lys Phe Tyr Ile Ile Gln Leu Leu Gln Asp Ser Asn Arg Phe Phe Thr $85 \ 90 \ 95$

Cys Trp Asn Arg Trp Gly Arg Val Gly Glu Val Gly Gln Ser Lys Ile 100 \$105\$

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Ala Gln Glu Ala Val Val Lys Val Asp Arg Gly Pro Val Arg Thr Val 165 170 175

Thr Lys Arg Val Gln Pro Cys Ser Leu Asp Pro Ala Thr Gln Lys Leu 180 185 190

Ile Thr Asn Ile Phe Ser Lys Glu Met Phe Lys Asn Thr Met Ala Leu 195 200 205

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Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu Ala Leu 225 230 235 240

Lys Gly Pro Thr Asp Gly Gly Gln Ser Leu Glu Glu Leu Ser Ser His 245 250 255

Phe Tyr Thr Val Ile Pro His Asn Phe Gly His Ser Gln Pro Pro

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Pro Thr Leu Gln His Ile Trp Lys Val Asn Gln Glu Glu Glu Asp 355 360 365

Arg Phe Gln Ala His Ser Lys Leu Gly Asn Arg Lys Leu Leu Trp His 370 375 380

Gly Thr Asn Met Ala Val Val Ala Ala Ile Leu Thr Ser Gly Leu Arg 385 390 395 400

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Glu Lys Arg Ile Ile Arg Val Asp Pro Thr Cys Pro Leu Ser Ser Asn

Pro Gly Thr Gln Val Tyr Glu Asp Tyr Asn Cys Thr Leu Asn Gln Thr

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Lys	Lys 290	Asp	Met	Leu	Leu	Val 295	Leu	Ala	Asp	Ile	Glu 300	Leu	Ala	Gln	Ala
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Thr	Gly	Ser 355	Asn	His	Arg	Cys	Pro 360	Thr	Leu	Gln	His	Ile 365	Trp	Lys	Va:
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	acc Thr										1173
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	ggc Gly										1461
	acc Thr										1509
	gtc Val										1557
	ctt Leu 485									ccg Pro	1605
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260 265 Asn Ser Pro Asp Val Leu Gln Ala Lys Lys Asp Met Leu Leu Val Leu 280 Ala Asp Ile Glu Leu Ala Gln Thr Leu Gln Ala Ala Pro Gly Glu Glu 295 Glu Glu Lys Val Glu Glu Val Pro His Pro Leu Asp Arg Asp Tyr Gln Leu Leu Arg Cys Gln Leu Gln Leu Leu Asp Ser Gly Glu Ser Glu Tyr Lys Ala Ile Gln Thr Tyr Leu Lys Gln Thr Gly Asn Ser Tyr Arg Cys Pro Asn Leu Arg His Val Trp Lys Val Asn Arg Glu Gly Glu Gly Asp Arg Phe Gln Ala His Ser Lys Leu Gly Asn Arg Arg Leu Leu Trp His Gly Thr Asn Val Ala Val Val Ala Ala Ile Leu Thr Ser Gly Leu Arg 390 Ile Met Pro His Ser Gly Gly Arg Val Gly Lys Gly Ile Tyr Phe Ala Ser Glu Asn Ser Lys Ser Ala Gly Tyr Val Thr Thr Met His Cys Gly Gly His Gln Val Gly Tyr Met Phe Leu Gly Glu Val Ala Leu Gly Lys Glu His His Ile Thr Ile Asp Asp Pro Ser Leu Lys Ser Pro Pro Pro Gly Phe Asp Ser Val Ile Ala Arg Gly Gln Thr Glu Pro Asp Pro Ala

Tyr Thr Val Ile Pro His Asn Phe Gly Arg Ser Arg Pro Pro Pro Ile

Gly Pro Pro Val Gln Cys Pro Ser Phe Lys Ser Ser Ser Phe Ser Gln 500 510

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e n v = v

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<222> (2)...(10), (12)...(13), (15)...(16), (20), (22)...(32)
<223> may be any amino acid; residue 32 may be present or absent
<400> 15
Leu Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Asn Xaa Xaa Tyr Xaa Xaa
                                    10
2.0
                                25
Trp Gly Arg Val Gly
        35
<210> 16
<211> 29
<212> PRT
<213> artificial sequence
<223> part-sequence motif 2
<220>
<221> VARIANT
<222>\ (2)\ldots(4)\,,\ (6)\,,\ (8)\ldots(11)\,,\ (14)\,,\ (16)\,,\ (18)\ldots(22)\,,\ (24)\ldots(26)\,,\ (28)
<223> may be any amino acid
<400> 16
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Ala Xaa Xaa Xaa Phe Xaa Lys Xaa Xaa Xaa Lys Thr Xaa Asn Xaa
Trp Xaa Xaa Xaa Xaa Phe Xaa Xaa Pro Xaa Lys
<210> 17
<211> 44
<212> PRT
<213> artificial sequence
<223> part-sequence motif 3
<220>
<221> VARIANT
<222> (2), (5)...(6), (8)...(16), (18)...(27), (33)...(35), (38)...(43)
<223> may be any amino acid
<220>
<221> VARIANT
<222> (4)
<223> Ile or Leu
<400> 17
Gln Xaa Leu Xaa Xaa Xaa Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
Met Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Aaa Pro Leu Gly Lys Leu
Xaa Xaa Xaa Gln Ile Xaa Xaa Xaa Xaa Xaa Leu
<210> 18
<211> 15
<212> PRT
<213> artificial sequence
<220>
<223> part-sequence motif 4
<220>
<221> VARIANT
<222> (4), (8), (11)...(13)
<223> may be any amino acid
<400> 18
Phe Tyr Thr Xaa Ile Pro His Xaa Phe Gly Xaa Xaa Xaa Pro Pro
                  5
                                    10
<210> 19
<211> 17
<212> PRT
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<213> artificial sequence
<223> part-sequence motif 5
<220>
<221> VARIANT
<222> (2)...(4), (6)...(7), (9), (13), (15)...(16)
<223> may be any amino acid
<400> 19
Lys Xaa Xaa Xaa Leu Xaa Xaa Leu Xaa Asp Ile Glu Xaa Ala Xaa Xaa
Leu
<210> 20
<211> 11
<212> PRT
<213> artificial sequence
<220>
<223> part-sequence motif 6
<220>
<221> VARIANT
<222> (2)...(4), (6)
<223> may be any amino acid
<400> 20
Gly Xaa Xaa Xaa Leu Xaa Glu Val Ala Leu Gly
                  5
<210> 21
<211> 28
<212> PRT
<213> artificial sequence
<223> part-sequence motif 7
<220>
<221> VARIANT
<222> (2)...(3), (5)...(8), (10)...(12), (14)...(22), (24), (26)...(27)
<223> may be any amino acid; residues 21 and 22 may be present or absent
<400> 21
Gly Xaa Xaa Ser Xaa Xaa Xaa Gly Xaa Xaa Pro Xaa Xaa Xaa
Xaa Xaa Xaa Xaa Xaa Leu Xaa Gly Xaa Xaa Val
                                 25
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<210> 22
<211> 16
<212> PRT
<213> artificial sequence
<220>
<223> part-sequence motif 8
<220>
<221> VARIANT
<222> (2)
<223> Tyr or Phe
<220>
<221> VARIANT
<222> (3)...(4), (6)...(8), (10)...(13)
<223> may be any amino acid
<400> 22
Glu Xaa Xaa Xaa Tyr Xaa Xaa Xaa Gln Xaa Xaa Xaa Tyr Leu Leu
<210> 23
<211> 20
<212> PRT
<213> artificial sequence
<223> synthetic sequence for antibody production
<400> 23
Met Ala Arg Arg Arg Arg Ser Thr Gly Gly Arg Ala Arg Ala
                                                         15
Leu Asn Glu Ser
<210> 24
<211> 20
<212> PRT
<213> artificial sequence
<223> synthetic sequence for antibody production
<400> 24
Lys Thr Glu Leu Gln Ser Pro Glu His Pro Leu Asp Gln His Tyr Arg
Asn Leu His Cys
             20
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<210> 25
<211> 21
<212> PRT
<213> artificial sequence
<223> synthetic sequence for antibody production
<400> 25
Cys Lys Gly Arg Gln Ala Gly Arg Glu Glu Asp Pro Phe Arg Ser Thr
Ala Glu Ala Leu Lys
             20
<210> 26
<211> 20
<212> PRT
<213> artificial sequence
<223> synthetic sequence for antibody production
<400> 26
Cys Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu
Glu Ala Leu Lys
<210> 27
<211> 19
<212> PRT
<213> artificial sequence
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<223> synthetic sequence for antibody production
<400> 27
Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu
Ala Leu Lys
<210> 28
<211> 19
<212> PRT
<213> Mus musculus
<400> 28
Lys Gln Gln Ile Ala Arg Gly Phe Glu Ala Leu Glu Ala Leu Glu Glu
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Ala Met Lys
<210> 29
<211> 7
<212> PRT
<213> artificial sequence
<220>
<223> NAD+ binding domain
<220>
<221> VARIANT
<222> (2)...(4)
<223> may be any amino acid residue
<400> 29
Gly Xaa Xaa Gly Lys Gly
<210> 30
<211> 38
<212> PRT
<213> Artificial Sequence
<223> PARP zinc finger sequence motif
<220>
<221> VARIANT
<222> (2)...(3), (5)...(34), (36)...(37)
<223> may be any amino acid; residues 33 and 34 may be present or absent
Xaa Xaa His Xaa Xaa Cys
<210> 31
<211> 10
<212> PRT
<213> Arabidopsis thaliana
<400> 31
Ala Ala Val Leu Asp Gln Trp Ile Pro Asp
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<210> 32